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(54) Title: CALCINEURIN INTERACTING PROTEIN COMPOSITIONS AND METHODS

(57) Abstract

identification characterization of a calcineurin interacting (CNI) protein effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin is described herein. One embodiment of the invention is the CNI polypeptide encoded by the CNI gene of Saccharomyces cerevisiae. Polynucleotides encoding a CNI protein are also described. Also described are yeast cells carrying mutations in the CNI gene. Further, a method of identifying a small molecule immunosuppressant compound is described. The methods include the use of a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains a subunit of calcineurin, and the other

Sau3A1 CNIC 1.22 kb Stop ATG Munl **BcoRV** CNI 3.5 kb ATG Stop

of two fusion hybrid proteins contains an immunophilin.

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CALCINEURIN INTERACTING PROTEIN COMPOSITIONS AND METHODS

Field of the Invention

The present invention relates to compounds affecting the function of calcineurin, particularly interactions of calcineurin with immunosuppressant drugs.

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30 Background of the Invention

The immune system functions as the body's major defense against diseases caused by invading organisms. This complex system fights disease by killing invaders such as bacteria, viruses, parasites or cancerous cells while leaving the body's normal tissues unharmed. The immune system's ability to distinguish the body's normal tissues, or self, from foreign or

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cancerous tissue, or non-self, is an essential feature of normal immune system function. A second essential feature is memory, the ability to remember a particular foreign invader and to mount an enhanced defensive response when the previously encountered invader returns. The loss of recognition of a particular tissue as self and the subsequent immune response directed against that tissue produce serious illness.

An autoimmune disease results from the immune system attacking the body's own organs or tissues, producing a clinical condition associated with the destruction of that tissue. An autoimmune attack directed against the joint lining tissue results in rheumatoid arthritis; an attack against the conducting fibers of the nervous system results in multiple sclerosis. The 10 autoimmune diseases most likely share a common pathogenesis and the need for safe and effective therapy. One type of therapy that has been employed in combating autoimmune disease is treatment with immunosuppressant drugs, such as cyclosporin A, FK506 and rapamycin. While the treatments are often effective, the drugs typically have undesirable side effects, including neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Many of these side effects are due to the drugs' action on cells other than those of the immune system.

In addition to their use in treating autoimmune conditions, immunosuppressive agents have also been used in treating or preventing transplantation rejection. Organ transplantation involving human organ donors and human recipients (allogeneic grafts), and non-human primate donors and human recipients (xenogeneic grafts), has received considerable medical and scientific attention (e.g., Roberts, 1989; Platt, 1990). To a great extent, this effort has been aimed at eliminating, or at least reducing, the problem of rejection of the transplanted organ. In the absence of adequate immunosuppressive therapy, the transplanted organ is destroyed by the host immune system.

Presently, the most commonly used agents for preventing transplant rejection include corticosteroids, cytotoxic drugs that specifically inhibit T cell activation such as azathioprine, immunosuppressive drugs such as cyclosporin A, and specific antibodies directed against T lymphocytes or surface receptors that mediate their activation (Briggs, 1991; Kennedy, 1983; Storb, 1985; Storb, 1986). All of these drug therapies are limited in effectiveness, in part because the doses needed for effective treatment of transplant rejection may increase the patient's susceptibility to infection by a variety of opportunistic invaders, and in part because of direct toxicity and other side effects.

Cyclosporin A, currently the most effective and most commonly used agent, is significantly toxic to the kidney. This nephrotoxicity limits the quantity of drug that can be

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safely given. The physician is frequently forced to administer sub-optimal doses of the drug because of this toxicity. A preparation capable of potentiating the action of immunosuppressive agents such as cyclosporin A on the immune system, thus allowing the administration of lower doses of drug, would be of considerable value in reducing the morbidity and mortality associated with transplantation.

Summary of the Invention

In one embodiment, the present invention includes polypeptide compositions effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin. The present invention includes the isolation and characterization of a calcineurin interacting protein, CNI, having these properties. Also disclosed herein are methods for the isolation and characterization of further CNI-related sequences and sequences of CNI-variants. The amino acid sequences presented as SEQ ID NO:2 and SEQ ID NO:5 are exemplary of the polypeptides of the present invention.

The present invention also includes a CNI polypeptide fragment that interacts specifically with the "A" subunits of calcineurin (CNAI and CNA2), but not with calcineurin "B" subunit (CNB1). In one embodiment, this fragment has an amino acid sequence of between 15 and 915 amino acids in length, for example, the c-terminal 306 amino acids of the CNI protein (CNIc).

Included aspects of the invention are an CNI polypeptide; a recombinant CNI polypeptide; and a fusion polypeptide comprised of an CNI polypeptide. Exemplary fusion proteins include fusions to β -galactosidase.

The invention further includes isolated nucleic acid sequences encoding the above described polypeptides and polypeptide fragments. Exemplary nucleic acid sequences include the sequences presented as SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:6. The present invention includes CNI-encoding genomic polynucleotides, cDNAs thereto and complements thereof. With respect to polynucleotides, some aspects of the invention include: a purified CNI-encoding genomic polynucleotide; CNI polypeptide-encoding RNA and DNA polynucleotides; recombinant CNI polypeptide-encoding polynucleotides; a recombinant vector including any of the above recombinant polynucleotides, and a host cell transformed with any of these vectors. Another aspect of the invention is a polynucleotide probe for CNI polypeptide-encoding sequences.

Portions of a CNI-polypeptide coding sequences are effective as probes to isolate variants coding sequences which occur naturally, or to determine the presence of such coding

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sequences in nucleic acid samples. Such probes include hybridization screening probes and polymerase chain reaction amplification primers specific for CNI-polypeptide coding sequences. Homologues of CNI may be isolated from a number of sources, such as other types of yeast cells (e.g., Schizosaccharomyces) or mammalian cells (e.g., human).

Other aspects of the invention include: a recombinant expression system which incorporates an open reading frame (ORF) derived from CNI polypeptide-encoding sequences, wherein the ORF is linked operably to a control sequence which is compatible with a desired host, a cell transformed with the recombinant expression system, and a polypeptide produced by the transformed cell. Typically the expression system includes a vector having (a) a nucleic acid containing an open reading frame that encodes a CNI-polypeptide; and (b) regulatory sequences effective to express said open reading frame in a host cell. The regulatory sequence may include sequences useful for targeting or secretion of the CNI-polypeptide: such as a secretory signal recognized in yeast or bacterial expression systems.

The invention includes a method of recombinantly producing CNI-polypeptides. In the method, a recombinant expression system containing an open reading frame (ORF) having a polynucleotide sequence which encodes a CNI-polypeptide, where the vector is designed to express the ORF in the host, is introduced into suitable host cells. The host is then cultured under conditions resulting in the expression of the ORF sequence. The CNI-polypeptide sequences discussed above are examples of suitable CNI-polypeptides. Numerous vectors and their corresponding hosts are useful in the practice of this method of the invention, including, but not limited, to the vectors described herein for expression in yeast cells, and lambda gt11 phage vector and E. coli cells. Other host cells include insect and mammalian cell expression systems.

The invention also includes purified antibodies that are immunoreactive with a CNI-polypeptide. The antibodies may be polyclonal or monoclonal. Antibodies that are specifically immunoreactive with CNI-polypeptides may be useful for the isolation of CNI-polypeptide homologues from other cell type sources (e.g., mammalian).

The present invention also includes, a method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed where one of two fusion hybrid proteins in the cell contains a subunit of calcineurin, and the other of two fusion hybrid proteins contains an immunophilin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. In one embodiment, the method is carried out using yeast cells, where one of the two

fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells (including human cells). The subunit may, for example, be yeast calcineurin subunit CNA1 or CNA2, or human calcineurin subunit "A". The immunophilin can, for example, be cyclophilins or FK506-binding proteins (e.g., FKBP12) typically from a homologous cell source.

Also included in the present invention is another method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed, wherein one of two fusion hybrid proteins in a cell contains an "A" subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI polypeptide. The cell preferably, but not necessarily, also contains a vector construct causing overexpression, or increased expression, of a "B" subunit of calcineurin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. This method is used to identify compounds (like FK506) that potentiate the interaction between CNI and CNA1. In one embodiment, the method is carried out using yeast cells, where one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells (including human cells). The subunits may, for example, be calcineurin subunit A1 or A2. The CNI polypeptide may also be from any source (e.g., yeast or human), and may be only a fragment of a complete CNI polypeptide (such as a c-terminal fragment). An exemplary cterminal fragment of CNI is CNIc.

Further, included in the present invention, is a yeast cell carrying a mutation in the naturally-occurring copy of CNI, where the mutation prevents expression of a functional CNI protein from the genomic copy. Embodiments of this aspect of the present invention include deletion mutations within the coding region of the CNI gene, deletion of regulation regions of the CNI gene, and non-sense or mis-sense mutations in the CNI gene. Yeast cells having such mutations are useful, for example, in a method of identifying proteins of similar function to CNI. In one embodiment, a hybrid interaction screen is set up in a cell with a CNI deletion and a GAL4 protein binding domain-CNA fusion and a GAL4 activation domain-immunophilin fusion. Expression libraries are then screened to identify clones encoding proteins that potentiate an interaction of an immunophilin with calcineurin. This screen will identify CNI-coding sequences as well as other proteins with a similar function.

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In a related embodiment, a yeast cell with a CNI deletion is used to identify CNI homologues (e.g., from other organisms, such as human) using a complementation assay or screen. Expression libraries (e.g., human lymphocyte expression libraries) are transformed into cells with a CNI deletion, and transformants are selected on their ability to complement the function of yeast CNI. An exemplary assay for selecting such transformants is exposure to hygromycin B. Cells which become more sensitive to hygromycin B following transformation are further analyzed to determine if the plasmid with which they were transformed contains an insert homologous to yeast CNI, or encoding a polypeptide with similar function to CNI.

The invention also includes a yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where the mutation prevents expression of a functional calcineurin-interacting polypeptide from the genomic copy. The mutation may be a null mutation, such as described in Example 8 below, or a different type of mutation, e.g., a nonsense or missense mutation. Nonsense and missense mutations may be generated using standard methods.

These and other objects and features of the invention will be more fully appreciated when the following detailed description of the invention is read in conjunction with the accompanying drawings.

Brief Description of the Figures

Figures 1A and 1B present schematic diagrams of sequences encoding the c-terminal portion of CNI (CNIc) fused to GAL-4 activation domain (GAL-4AD) (Fig. 1A), and sequences encoding CNI (Fig. 1B).

Figure 2A presents data from a β -galactosidase (β -gal) assay to detect the interaction of CNIc with the A1 subunit of calcineurin (CNA1), A2 subunit of calcineurin (CNA2), GAL-4 binding domain (G4BD) and lamin C. A labeled schematic diagram corresponding to the data shown in Fig. 2A is presented in Fig. 2B to facilitate reference to individual groups of colonies.

Figure 3A presents data from a β -gal assay to detect the interaction of CNIc with CNA1 Δ C, CNA2 Δ C and CNB1. A labeled schematic diagram corresponding to the data shown in Fig. 3A is presented in Fig. 3B.

Figures 4A and 4C present data from β -gal assays to evaluate the effects of FK506 and the deletion of CNB1 on the interactions of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 4A and 4C are presented in Figs. 4B and 4D, respectively.

Figures 5A and 5C present data from β -gal assays to evaluate the effects of FK506 and the overexpression of CNB1 on the interactions of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 5A and 5C are presented in Figs. 5B and 5D, respectively.

Figures 6A, 6C and 6E present data from β -gal assays to evaluate the effects of FK506, cyclosporin A (CsA), and the deletion of CNB1 on the interaction of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 6A, 6C and 6E are presented in Figs. 6B, 6D and 6F, respectively.

Figures 7A, 7C, 7E and 7G present data from β-gal assays to evaluate the effects of FK506, cyclosporin A (CsA), rapamycin and the overexpression of CNB1 on the interaction of CNIc with CNA and CNAΔC. Labeled schematic diagrams corresponding to the data shown in Figs. 7A, 7C, 7E and 7G are presented in Figs. 7B, 7D, 7F and 7H, respectively.

Figure 8A presents data from a β -gal assay to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNA. A labeled schematic diagram corresponding to the data shown in Fig. 8A is presented in Fig. 8B.

Figures 9A and 9C present data from β -gal assays to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNB1. Labeled schematic diagrams corresponding to the data shown in Figs. 9A and 9C are presented in Figs. 9B and 9D, respectively.

Figure 10 presents an image of a protein blot of CNIc and CNA co-immunoprecipitate probed with anti-CNA2 antibody.

Figure 11 presents an image of a yeast RNA blot hybridized with a CNIc probe.

Figure 12 presents the DNA sequence of a 3.5 kb fragment of yeast chromosome 11 containing the coding sequence for a yeast CNI protein.

25 Figure 13 presents the amino acid sequence of a yeast CNI protein.

Brief Description of the Sequences

SEQ ID NO:1 presents the nucleotide sequence of a Sau3AI fragment containing the coding sequence for CNIc.

SEQ ID NO:2 presents the amino acid sequence of CNIc encoded by SEQ ID NO:1. SEQ ID NO:3 presents the coding sequence presented in SEQ ID NO:1.

SEQ 1D NO:4 presents the nucleotide sequence of a gene encoding a complete CNI protein.

SEQ ID NO:5 presents the amino acid sequence encoded by SEQ ID NO:4.

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SEQ ID NO:6 presents the coding sequence presented in SEQ ID NO:4.

SEQ ID NO:7 presents the nucleotide sequence of PCR primer CNI-PCR-A.

SEQ ID NO:8 presents the nucleotide sequence of PCR primer CNI-PCR-B.

SEQ ID NO:9 presents the nucleotide sequence of a gene encoding the yeast CNA1 subunit of calcineurin.

SEQ ID NO:10 presents the amino acid sequence encoded by SEQ ID NO:9.

SEQ ID NO:11 presents the nucleotide sequence of a gene encoding the yeast CNA2 subunit of calcineurin.

SEQ ID NO:12 presents the amino acid sequence encoded by SEQ ID NO:11.

SEQ ID NO:13 presents the nucleotide sequence of a gene encoding the yeast CNB1 subunit of calcineurin.

SEQ ID NO:14 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:15 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:16 presents the coding sequence presented in SEQ ID NO:13.

SEQ ID NO:17 presents the amino acid sequence encoded by SEQ ID NO:16.

SEQ 1D NO:18 presents a nucleotide sequence encoding CNA1\(Delta c.\)

SEQ ID NO:19 presents the amino acid sequence encoded by SEQ ID NO:18.

SEQ 1D NO:20 presents a nucleotide sequence encoding CNA2Δc.

SEQ ID NO:21 presents the amino acid sequence encoded by SEQ ID NO:20.

SEQ ID NO:22 presents the nucleotide sequence of PCR primer G4-PCR-A.

SEQ ID NO:23 presents the nucleotide sequence of PCR primer G4-PCR-B.

Detailed Description of the Invention

I. DEFINITIONS

A "calcineurin-targeted immunosuppressant" is a compound that possesses in vivo immunosuppressive activity, and that interacts with an immunophilin to form a complex which is capable of inhibiting calcineurin.

"Interacting proteins" are proteins capable of specifically binding to one another, or associating with one another, in a cell or in vitro.

A calcineurin interacting (CNI) protein or polypeptide is a protein or polypeptide that is effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin. Preferably, a CNI protein or polypeptide is a protein or polypeptide having an amino acid sequence that is homologous to the sequence presented herein as SEO ID NO:5.

"Substantially isolated" is used in several contexts and typically refers to the at least partial purification of a CNI protein or polypeptide fragment away from unrelated or contaminating components (ϵ .g., cytoplasmic contaminants and heterologous proteins). Methods and procedures for the isolation or purification of compounds or components of interest are described below (ϵ .g., affinity purification of fusion proteins and recombinant production of CNI polypeptides).

In the context of the present invention, the phrase "nucleic acid sequences," when referring to sequences which encode a protein, polypeptide, or peptide, is meant to include degenerative nucleic acid sequences which encode homologous protein, polypeptide or peptide sequences as well as the disclosed sequence.

Two nucleic acid fragments are considered to have "homologous" sequences if they are capable of hybridizing to one another (i) under typical hybridization and wash conditions, as described, for example, in Sambrook, et al., pages 320-328, and 382-389, or (ii) using reduced stringency wash conditions that allow at most about 25-30% basepair mismatches, for example:

2 × SSC, 0.1% SDS, room temperature twice, 30 minutes each; then 2 × SSC, 0.1% SDS, 37°C once, 30 minutes; then 2 × SSC, room temperature twice, 10 minutes each. Preferably, homologous nucleic acid strands contain 15-25% basepair mismatches, even more preferably 5-15% basepair mismatches. These degrees of homology can be selected by using wash conditions of appropriate stringency for identification of clones from gene libraries (or other sources of genetic material), as is well known in the art.

Two amino acid sequences or two nucleotide sequences (in an alternative definition for homology between two nucleotide sequences) are considered homologous (as this term is preferably used in this specification) if they have an alignment score of >5 (in standard deviation units) using the program ALIGN, typically default mutation gap matrix and gap penalty (Dayhoff). The two sequences (or parts thereof) are more preferably homologous if their amino acids are greater than or equal to 40% using the ALIGN program mentioned above.

II. OVERVIEW OF INVENTION

Experiments performed in support of the present invention demonstrate the identification and isolation of the nucleic acid sequence encoding a calcineurin interacting (CNI) protein. Further experiments performed in support of the present invention characterize the CNI protein, as well as a polypeptide containing only the c-terminal 306 amino acids of the CNI protein (CNIc). The experiments demonstrate that CNIc interacts specifically with the "A" subunits of calcineurin (CNA1 and CNA2), but not with calcineurin "B" subunit (CNB1).

The experiments also demonstrate that CNIc does not interact directly with FK506 binding protein (FKBP; Schreiber, et al.; with or without FK506), GAL4 binding domain (G4_{BD}) or lamin C. The experiments also demonstrate that CNIc interacts with C-terminally truncated forms of CNA (CNA Δ C), which have lost their autoinhibitory domains, though the interaction is somewhat weaker that with full length CNA proteins.

Additional experiments show that the interaction between CNIc and CNA is enhanced when CNB1 is deleted, and diminished when CNB1 is overexpressed, that the interaction between CNIc and CNA or CNA\(Delta\)C is markedly enhanced by FK506 and by Cyclosporin A (CsA), but not rapamycin, and that overexpression of a full-length CNI protein enhances the interaction between CNA and FKBP (detectable only in the presence of FK506).

Additional experiments conducted in support of the present invention demonstrate that overexpression of the full-length CNI has no detectable effect on the interaction between CNB1 and CNA, and that in the presence of FK506 or CsA, overexpression of CNB1 no longer inhibits the interaction of CNIc with CNA.

It was also found that CNI deletion mutants are viable, both in wild-type and CN-deletion backgrounds, and that CNI deletion mutants in a CN-deletion background are more resistant to hygromycin B than normal CN-deletion mutants.

Co-immunoprecipitation experiments demonstrate that CNIc and CNA co-immunoprecipitate in the presence of FK506, and protein blot experiments show that CNI is expressed at low levels *in vivo*. RNA blot experiments show that CNI is encoded by a single message approximately 2.9 kb in length.

A comparison of the yeast CNI sequence with sequences present in nucleic acid and amino acid databases reveals no obvious homologous sequences have been identified in other organisms.

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III. CALCINEURIN

Experiments performed in support of the present invention were designed to identify polypeptides capable of interacting with calcineurin. Calcineurin (also called phosphoprotein phosphatase 2B or PP2B), has been characterized from many different tissues and organisms (Klee, et al.). It is a heterodimer of two subunits, of which the "A" subunit is about 61 kD in weight, possesses catalytic activity and also contains the association site for calmodulin. The "B" subunit contains four Ca2+ binding sites and activates the A subunit. Calcineurin has little enzymatic activity, even in the presence of Ca2+ and only becomes fully active when associated with calmodulin (Cyert).

Two A subunits (CNA1 and CNA2; Cyert, et al., 1991) and one B subunit (CNB1, Cyert, et al., 1992) have been cloned in yeast. Either CNA1 or CNA2 may associate with CNB1 to form a functional calcineurin heterodimer. Multiple isotypes of the A subunit have been cloned from a variety of organisms and are highly conserved (Klee, et al.). In particular, calcineurin subunits have been cloned from human tissue (see reviews by, for example, Klee, et al., and Guerini, et al.).

IV. IMMUNOSUPPRESSANT DRUGS

FK506, cyclosporin A (CsA) and rapamycin, derived from fungi, inhibit the activation of T-cells by antigens. The compounds have proven highly effective at suppressing mammalian immune systems in vivo. In particular, CsA therapy in clinical settings has dramatically increased the success rate of transplantation therapy.

It is now known that FK506 and CsA exert their immunosuppressive effects, in part, by inhibiting the transcriptional activation of the interleukin-2 (IL-2) gene, whereas rapamycin appears to function by inhibiting the response of T-cells to IL-2, presumably by inhibiting a transduction pathway mediated by the IL-2 receptor.

The molecular mechanism of FK506 and CsA immunosuppressive action involves a group of small, abundant intracellular proteins termed immunophilins, which bind with a high affinity to the immunosuppressants (Schreiber). At least two classes of immunophilins are known to exist. One class, termed cyclophilins, binds to CsA, while another class, the FK506-binding proteins (FKBPs) binds FK506 and rapamycin. Many immunophilin genes, from a variety of organisms, have been cloned, and appear to be highly conserved from simple eukaryotes to mammals.

It is believed that FK506 and CsA-induced immunosuppression is due to the binding of complexes, formed by binding of immunosuppressants FK506 and CsA bound to one of their respective immunophilins, to the catalytic subunit of calcineurin (Schreiber, et al., Liu, et al., Foor, et al., Weiss, et al.). The binding of such a complex to an (A) subunit inhibits activation of calcineurin by increased intracellular calcium, which in turn prevents calcineurin from activating transcription factor NF-AT. Since IL-2 is one of the genes controlled by NF-AT in T-cells, inhibition of the transcription factor inhibits the production of IL-2, resulting in immunosuppression (Clipstone, et al.).

FK506 and CsA are widely used in organ transplantation to prevent host rejection. However, both drugs are known to have many undesired side-effects such as neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Accordingly formulations effective to

increase a target cell's sensitivity to these drugs may be useful in alleviating some of the aforementioned side-effects. Specifically, CNI and its homologues or derivatives, administered at appropriate levels, may be able to increase the sensitivity of CN to FK506/CsA and reduce the necessary dosage thus reducing or eliminating the side-effects of these drugs.

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V. Two Hybrid Protein Interaction Assays

Two hybrid protein interaction assay methods (two hybrid protein-protein interaction screens) provide a simple and sensitive means to detect the interaction between two proteins in living cells. The assays are based on the finding that most eukaryotic transcription activators are modular (e.g, Brent, et al.), i.e., that the activators typically contain activation domains that activate transcription, and DNA binding domains that localize the activator to the appropriate region of a DNA molecule.

The development of two hybrid protein interaction assays was made possible by the observation that the DNA binding domain does not need to be physically located on the same polypeptide as the activation domain (Ma, et al., Triezenberg, et al.), raising the possibility that transcription of reporter genes could be used as an assay to detect protein interactions.

The utility of two hybrid systems for detecting interactions between two interacting proteins was fully realized by the observation that protein interactions could be detected if two potentially-interacting proteins were expressed as fusions, or chimeras (Fields, et al.). A first fusion protein contains one of a pair of interacting proteins fused to a DNA binding domain, and a second fusion protein contains the other of a pair of interacting proteins fused to a transcription activation domain. The two fusion proteins are independently expressed in the same cell, and interaction between the "interacting protein" portions of the fusions reconstitute the function of the transcription activation factor, which is detected by activation of transcription of a reporter gene.

At least two different cell-based two hybrid protein-protein interaction assay systems have been used to assess binding interactions and/or to identify interacting proteins. Both employ a pair of fusion hybrid proteins, where one of the pair contains a first of two "interacting" proteins fused to a transcription activation domain of a transcription activating factor, and the other of the pair contains a second of two "interacting" proteins fused to a DNA binding domain of a transcription activating factor.

The yeast GAL4 two hybrid system (Fields, et al., Chien, et al., Durfee, et al., Bartel, et al.), utilized for experiments performed in support of the present invention, was developed

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to detect protein-protein interaction based on the reconstitution of function of GAL4, a transcriptional activator from yeast, by activation of a GAL1-lacZ reporter gene.

Like several other transcription activating factors, GAL4 contains two distinct domains, a DNA binding domain and a transcription activation domain. Each domain can be independently expressed as a portion of a fusion protein composed of the domain, and a second, "bait" interacting protein. The two fusion proteins are then independently expressed together in a cell. When the two GAL4 domains are brought together by a binding interaction between the two "interacting" proteins, transcription of a reporter gene under the transcriptional control of GAL4 is initiated. The reporter gene typically has a promoter containing GAL4 protein binding sites (GAL upstream activating sequences, UAS₀). Exemplary reporter genes are the GAL1-lacZ, and GAL1-HIS3 reporter genes used in experiments described herein.

A second two hybrid system, described in detail in Ausubel, et al., utilizes a native E. coli LexA repressor protein, which binds tightly to appropriate operators. A plasmid is used to express one of a pair of interacting proteins (the "bait" protein) as a fusion to LexA. The plasmid expressing the LexA-fused bait protein is used to transform a reporter strain of yeast, such as EGY48, that contains pSH18-34.

In this strain, binding sites for LexA are located upstream of two reporter genes. In the first reporter system, the upstream activation sequences of the chromosomal LEU2 generequired in the biosynthetic pathway for leucine (Leu)—are replaced in EGY48 with lexA operators, permitting selection for viability when cells are plated on medium lacking Leu. In the second reporter system, EGY48 harbors a plasmid, pSH18-34, that contains a lexA operator-lacZ fusion gene, permitting discrimination based on color when the yeast is grown on medium containing Xgal (Ausubel, et al.).

25 promoter to express proteins as fusions to an acidic domain ("acid blob") that functions as a portable transcriptional activation motif ("act"), and to other useful moieties. Expression of library-encoded proteins is induced by plating transformants on medium containing galactose (Gal), so yeast cells containing library proteins that do not interact specifically with the bait protein fail to grow in the absence of Leu. Yeast cells containing library proteins that interact with the bait protein form colonies within 2 to 5 days, and the colonies turn blue when the cells are streaked on medium containing Xgal. The plasmids are isolated and characterized by a series of tests to confirm specificity of the interaction with the initial bait protein. Those found to be specific are ready for further analysis (e.g., sequencing).

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LexA and GAL4 each have different properties that should be considered when selecting a system. LexA is derived from a heterologous organism, has no known effect on the growth of yeast, possesses no residual transcriptional activity, can be used in GAL4* yeast, and can be used with a Gal-inducible promoter. Because GAL4 is an important yeast transcriptional activator, experiments must be performed in gal4 yeast strains to avoid background from endogenous GAL4 activating the reporter system.

Both two hybrid systems have been successfully used for isolating genes encoding proteins that bind a target protein and as simple protein binding assays (e.g., Yang, et al., Gyuris, et al.), and both can be applied to methods of the present invention.

Both gene isolation and protein binding assay applications of the GALA system are described in Examples below.

VI. SPECIFIC EMBODIMENTS

Example 1 demonstrates application of an exemplary two hybrid protein-protein interaction screen (Materials and Methods, section D) to the screening of three pGAD yeast fusion libraries, carrying fusions between the transcription activating domain of yeast protein GAL4 (G4AD) and yeast genomic DNA Sav3Al fragments, in all three reading frames. The libraries are screened to identify polypeptides, encoded by the Sau3Al fragments, capable of interacting with catalytic (A) subunits of calcineurin, expressed as fusions with the GAL4 protein binding domain (GBT-CNA fusions).

Three sets of yeast cells harboring pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid and a GAL4-activated LacZ reporter gene are each transformed with one of the three reading-frame libraries. Construction of the plasmids used is described in Materials and Methods, sections B and C. Cells transformed with a plasmid encoding a protein fusion capable of interacting with the CNA subunit fusion are selected using a β -galactosidase (β -gal) assay on plates containing the chromogenic substrate X-gal (Materials and Methods, section E). Results of the β -gal assay are confirmed using a growth assay (Materials and Methods, section F). False positives are eliminated by colony purification (re-streaking for single colonies), PCR experiments using GAL4 primers, and testing against a number of test fusions by β -gal assays on transformed haploid or mated diploid reporter strains.

A yeast clone encoding a polypeptide capable of specifically interacting with CNA polypeptide fusions is identified and sequenced. The sequence of the Sau3AI fragment is presented as SEQ ID NO:1. The coding sequence forming the open reading frame is presented as SEQ ID NO:3. The polypeptide encoded by the open reading frame is presented as SEQ

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ID NO:2. The open reading frame encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GAL4, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product. The clone is termed CNIc, with the lowercase "c" representing "c-terminal".

Figure 1A shows a schematic representation of the nucleic acid sequence encoding the GAL4AD-CNIc fusion protein. The stippled portion between GAL4AD and CNIc represents a linker discussed in the Materials and Methods section, as well as in Example 1. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons, and Sau3Al restriction sites.

Example 1 further describes the identification of a λ clone encoding a full length sequence version of CNIc, termed CNI. The polypeptide encoded by the sequence is termed CNI protein. The clone is identified by hybridization screening of a panel of λ clones spanning the yeast genome using a 1.22 kb ³²P-labeled probe generated from CNIc.

Phage lysates of the λ clone are amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments. A 3.16 kb MunI/EcoRV fragment from the λ clone insert contains the coding sequence of CNI. The sequence of the 3.16 kb MunI/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4, and a schematic diagram of the sequence is shown in Figure 1B. This sequence contains the entire 2.75 kb coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

A search of known DNA and protein sequences turns up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin interacting protein.

The methods referred to above may also be applied to the screening of, for example, a human cDNA library using an appropriate two-hybrid protein interaction screen. The "bait" protein in the interaction screen (e.g., the protein analogous to CNA1 in Example 1) may be of yeast origin (e.g., CNA1), but is preferably of human origin (e.g., a human calcineurin "A" subunit; Klee, et al.). The bait protein is expressed in the cell (e.g., a yeast cell) used for the two hybrid interaction screen as a fusion to a domain of a transcription activating factor (e.g., the DNA binding domain of GAL4). The library may be a human DNA library in a vector (e.g., pGAD) effective to express library sequences as fusions to a complimentary domain of the transcription activating factor (e.g., the activation domain of GAL4). Libraries of human sequences can be derived from a number of sources including genomic DNA, such as yeast

artificial chromosome (YAC) constructs carrying genomic human DNA, or cDNA generated from a variety of cell types (e.g., activated T-cells).

Example 2 details a β -gal assay to determine the specificity of binding of CNIc to subunits of calcineurin. Exemplary results are shown in Figure 2A. The legend for Fig. 2A is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs (indicated in Example 2).

A comparison of the intensities of the blue β -gal reaction product indicates that CNIc interacts strongly with CNA1 (21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with GAD-CNIc (26) show a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact with each other are used as a positive control for the assay (20). The data presented in Figure 2A show that CNIc interacts specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments, illustrated in Fig. 3A, is conducted using constitutivelyactive CNA subunits, as well as calcineurin subunit CNB1. As described in the Materials and Methods section below, CNA1ΔC and CNA2ΔC are each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A. The legend for Fig. 3A is shown in Figure 3B. The data show that GBT-A1\(\triangle\) and GAD-CNIc (28) gives a definite positive signal, while GBT-A2\(\triangle\) and 20 GAD-CNIc (29) is weaker, though still detectable above its background (i.e. GBT-A2ΔC and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) is not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 Δ C and GAD-B1 (34) and GBT-A2\(\Delta\) and GAD-B1 (35) give strong signals. The data presented in Figure 3A show that CNIc interacts specifically with CNA1 Δ C and CNA2 Δ C, but not with CNB1.

Example 3 details the effects of immunosuppressant drugs on binding of CNIc to calcineurin in B1st, B1 Deletion and B1 Overproducing Yeast Strains. The yeast strains are assayed for β -gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments are performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 30 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data are shown in Figures 4A, 4C, 5A and 5C. The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Figs. 4A and 5A illustrate experiment

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performed without FK506, while experiments shown in Figs. 4C and 5C were performed with FK506.

The interactions of various combinations of proteins expressed by constructs indicated in Example 3 was studied in three yeast strains, one of which is null for the CNB1 subunit of calcineurin (Y153b; at 36-40), while the others (Y190; at 41-46 and Y526 at 47-51) are wild-type for CNA1, CNA2 and CNB1.

The data, shown in Fig. 4A (no added drugs), illustrate that deleting the endogenous host CNB1 gene potentiates, or enhances, interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1 Δ C and CNA2 Δ C. Comparison of corresponding colonies in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA Δ C interactions. The drug enhances interactions under all except control conditions. The effect is most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhances, or potentiates CNIc-CNA/CNA\(Delta\)C interactions under conditions where the CNB1 subunit is overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNA\(Delta\)C interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) have reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhances, or potentiates interactions in all colonies, except the negative controls (56).

The data presented in Figures 4A, 4C, 5A and 5C demonstrate that the interaction of CNIc with CNA and CNA\(Delta\)C is markedly enhanced by FK506. The interaction is also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction is overcome by the stimulatory effect of FK506.

Stated another way, inclusion of a small molecule immunosuppressant (FK506) potentiates an interaction between two fusion hybrid proteins, where one of the two proteins contains an (A) subunit of calcineurin, and the other protein contains a CNI polypeptide. The potentiation is particularly strong when the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell (e.g., the expresses B1/YEp352).

In the present case, a yeast cell is modified to cause overexpression of a "B" subunit of calcineurin (CNB1) by transforming the cell with B1/YEp352 (construction described below). A cell may be modified to cause overexpression of a "B" subunit of calcineurin in other ways as well, such as, for example, transformation with other types of expression vectors encoding a "B" subunit of calcineurin, or treatment with a substance that upregulates a promoter controlling expression of an endogenous (B) subunit of calcineurin.

In light of the effects of FK506 on CNIc-CNA/CNA Δ C interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A is also effective to enhance interaction of CNIc with CNA and CNA Δ C. Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through calcineurin (Cyert).

Similarly, data shown in Figures 7A and 7C support data in Figures 5A and 5C, and further, demonstrate that there is no detectable interaction between FK506 binding protein (FKBP) and CNIc. Results shown in Fig. 7E demonstrate that cyclosporin A has a similar effect to FK506 in cells overexpressing CNB1 — that is, it enhances the interactions between CNIc and CNA/CNAAC.

In contrast, data presented in Fig. 7G show that the immunosuppressant rapamycin has no detectable effect on CNIc-CNA/CNA\(Delta\)C interactions (compare Fig. 7G with Fig. 7A).

Taken together, the data presented in Figs. 6A, 6C, 6E 7A, 7C, 7E and 7G show that like FK506, cyclosporin A (CsA), but not rapamycin, enhances the interaction of CNIc with CNA and CNAΔC, and that CNIc doesn't interact with FKBP with or without FK506.

Example 4 describes experiments to assess effects of CNIc on FKBP/FK506 binding to calcineurin. Figure 8A presents exemplary data from studies to assess the effect of CNI on FKBP-mediated FK506 interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 show no detectable interaction (84). In the presence of FK506, however, the proteins interact (85), presumably because FK506 forms a complex with FKBP, which then binds CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI has no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiates, or enhances the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

Experiments illustrated in Figures 9A and 9B demonstrate that FK506 has little or no effect on the binding of CNA to CNB1. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively. The data in Fig. 9A show that overexpression of the full-length CNI

clone markedly enhances the FK506-dependent interaction of FKBP with CNA, although it doesn't affect the interaction between CNA and CNB1.

Example 5 presents co-immunoprecipitation of CNIc (carrying an HA epitope tag) and CNA. Immunoprecipitation is carried out with anti-HA monoclonal antibody and the immune complex, resolved by SDS-PAGE, is detected with anti-CNA2 polyclonal antibody and visualized with goat anti-rabbit antibody using the "ECL" method (Amersham, Arlington Heights, IL). The results, shown in Figure 10, demonstrate that CNIc is capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. Similar methods may be employed to isolate a CNI analog from other cell sources, including mammalian (specifically human).

Example 6 describes yeast RNA blots hybridized with a CNIc probe. Exemplary data are shown in Figure 11. A single message of approximately 2.9 kb is detected. The data indicate that CNI is an expressed gene encoding a 2.9 kb message in yeast.

Example 8 details the construction of cni null mutants. The null mutants are employed to assess if CNI is required for viability in yeast, and to test hygromycin B sensitivity. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains can survive, but that CNI deletions render a host more resistant to hygromycin B. The effect is particularly pronounced in both MCY300-1 (cna1 cna2) and DD12 (cnb1), suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B.

20 It will be understood that all of the above methods and experimental manipulations are amenable to being done with interacting polypeptides from organisms other than yeast. In particular, calcineurin subunits, CNI polypeptides, immunophilins and the like may be of mammalian origin, e.g., human origin.

25 VII. <u>UTILITY</u>

Methods and compositions of the present invention may be applied in a number of different ways. Following the guidance presented herein, one of skill in the art may isolate nucleic acids encoding additional CNI polypeptides, for example, a human CNI polypeptide.

In one approach, a yeast strain carrying a mutation of the CNI gene, e.g., a deletion, is used to clone heterologous sequences (e.g., human sequences) by complementation. A library of genomic DNA or, preferably, cDNA from an organism (e.g., human) and tissue (e.g., lymphocyte cells) of choice is cloned into a vector that can be maintained in yeast. Preferably, the vector contains a yeast promoter effective to express the heterologous sequences in yeast cells. Several heterologous libraries suitable for expression in Saccharomyces

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cerevisiae containing DNA from S. pombe (Beach, et al.) and Drosophila have been constructed.

The library is transformed into a suitable yeast strain carrying a *cni* mutation, and transformants are selected using a suitable complementation assay. For example, transformants may be screened for increased hygromycin sensitivity, as experiments described herein indicate that *cni* deletion mutants possess a decreased sensitivity to hygromycin B (Example 8). The screen may be made more effective by using a yeast strain that is hypersensitive to hygromycin B, such as a strain deficient for a subunit of calcineurin (Example 8).

Alternatively, human CNI DNA sequences may be isolated by directly screening a library, e.g., a lymphocyte cDNA library, for clones hybridizing with a yeast CNI nucleic acid probe. The generation of an exemplary yeast CNI nucleic acid probe is described in Example 1.

In another approach, particularly advantageous for isolating sequences expressed at low levels, a CNI nucleic acid probe may be used to screen a genomic library, e.g., a human genomic library, to isolate a sequence that may be used to design probes or primers that may match the target sequence better that the yeast sequence. Such primers may be used with, for example, PCR, to isolate longer fragments from a tissue-specific library.

In yet another approach, an antibody generated against CNI polypeptide is used to immunoprecipitate a CNI polypeptide from an organism and/or tissue of choice. The protein may then be micro-sequenced, and the sequence utilized to design degenerate primers useful for isolating a cDNA.

CNI polypeptides of the present invention, particularly CNI fragments that retain a desired binding activity, may be used as lead compounds useful for the development of small molecules having cellular functions similar to those of the CNI-polypeptides, that is, molecules effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin.

CNI-polypeptides of the present invention may also be employed in a method of increasing sensitivity of cells to calcineurin-affecting immunosuppressant drugs. In this method, a CNI-polypeptide is introduced into the cell typically prior to or at the same time as contacting the cell with an immunosuppressant drug, such as FK506. The polypeptide may be delivered by any suitable means effective to deliver polypeptides to selected cells.

Alternatively, nucleic acids encoding CNI polypeptides may be used in appropriate expression vectors as a genetic therapy tool to potentiate the immunosuppressive effects of

calcineurin-targeting immunosuppressant drugs. The vectors may be targeted to selected cells, such as T-cells, to increase their sensitivity to a given systemic dose of an immunosuppressant.

Another utility of the present invention includes methods of screening for substances that up-regulate expression of CNI polypeptides, *i.e.*, substances that affect transcription. Such substances are useful for sensitizing cells to immunosuppressant drugs. In this method, the CNI promoter can be attached to a gene that functions as a selectable marker (for use in genetic selections to screen test substances) or to a reporter gene (for use in evaluating the effect on CNI transcription by test substances).

In another aspect of the present invention, the CNI-polypeptides, for example, mammalian homologue polypeptides of CNI, have potential use as therapeutic agents for both human and veterinary use. For example, CNI-polypeptides may be used in a method of enhancing immunosuppression in a test subject. In this method, the CNI-polypeptide is administered to the subject in a pharmaceutically-acceptable formulation and at a concentration effective to potentiate the interaction of an immunosuppressant/immunophilin complex with a subunit of calcineurin. The method may also include contacting the CNI-polypeptide with a cell under conditions effective to permit uptake of the protein into the cell in order to increase sensitivity of the cell to immunosuppressants. A CNI polypeptide used in such methods may be modified to be more suitable for administration or to be more effective in a cell. For example, a CNI polypeptide may be modified to eliminate PEST motifs, which are typically found in proteins with short half-lives, to extend the effective lifetime of the polypeptide in the target cell.

The following examples illustrate, but in no way are intended to limit the present invention.

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MATERIALS AND METHODS

Unless indicated otherwise, chemicals and reagents were obtained from Sigma Chemical Company (St. Louis, MO) or Mallinckrodt Specialty Chemicals (Chesterfield, MO), restriction endonucleases were obtained from New England Biolabs (Beverly, MA), and other modifying enzymes and biochemicals were obtained from Pharmacia Biotech (Piscataway, NJ) or Boehringer Mannheim (Indianapolis, IN). FK506 was obtained from Fujisawa USA, Inc. (Deerfield, IL), cyclosporin A was obtained from Sandoz (Basel, Switzerland), and rapamycin was obtained from Wyeth-Ayerst (Princeton, NJ). Materials for media for yeast growth and culture were obtained from DIFCO (Detroit, MI). Unless otherwise indicated, manipulations

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of yeast, bacteria, nucleic acids, proteins and antibodies were performed using standard methods and protocols (e.g., Guthrie, et al., Sambrook, et al., Ausubel, et al., Harlow, et al., and Rose, et al.).

5 A. Buffers

Z buffer: 60 mM Na₂HPO₄-7H₂O, 40 mM NaH₂PO₄-H₂O, 10 mM KCl, 1 mM MgSO₄-7H₂O and 50 mM β -mercaptoethanol (pH 7.0).

B. Plasmids, Libraries and Yeast Strains

Plasmids pGBT9 (GBT), carrying GAL4 DNA-binding domain (amino acid residues 1-147; G4BD) and TRP1, and pGAD (GAD), carrying GAL4 activation domain (amino acid residues 768-881; G4AD) and LEU2; three pGAD libraries carrying fusions between G4AD and yeast genomic Sau3AI partial-digest fragments in each frame; and the yeast GAL1-lacZ reporter strain SFY526 (Y526; MATa ura3-52 ade2-101 lys2-801 his3-200 urp1-901 leu2-3,112 can1 gal4-542 gal80-538 URA3::GAL1-lacZ) were obtained from Stanley Fields (State University of New York at Stony Brook, Stony Brook, NY; Chien, et al., Bartel, et al.). The libraries were constructed with linkers between the GAL4 activation domain and the Sau3AI fragments. The sequences of the linkers were 5'-ATCG-3'for the first library, 5'-ATCCG-3'for the second library, and 5'-ATCCCG-3'for the third library. In this way, the yeast genomic Sau3AI fragments were cloned in all three reading frames relative to G4AD.

Plasmids pAS2 (AS) carrying G4BD and TRP1, and pAS-lamin (AS-lamin) containing a sequence encoding a G4BD-lamin C fusion; and yeast reporter strains Y190 (MATa ura3-52 ade2-101 his3-\(\Delta\)200 trp1-901 leu2-3,112 cyh2\(\Delta\), gal4\(\Delta\) gal80\(\Delta\) URA3::GAL-lacZ LYS2::GAL-HIS3), a derivative of Y153 carrying dual indicator genes (GAL-lacZ and GAL-HIS3), and Y187 (MAT\(\Omega\) ura3-52 ade2-101 lys2-801 his3-200 trp1-901 leu2-3,112 gal4\(\Delta\) gal80\(\Delta\) URA3::GAL-lacZ) carrying GAL-lacZ reporter were obtained from Stephen Elledge (Baylor College of Medicine, Houston, TX; Durfee, et al.). Yeast strain Y153b1 (cnb1::ADE2)was derived from Y153.

E coli strain JBe181 (leuB600 trpC9830) was obtained from Ira Herskowitz (University of California at San Francisco, San Francisco, CA). Protease-deficient yeast strain BJ2407 (Guthrie, et al.) was obtained from the Yeast Genetic Center (University of California at Berkeley, Berkeley, CA).

C. **GAL4-Calcineurin Fusions**

GAL4-calcineurin (GAL4-CN) fusions, GBT-A1 (G4BD-CNA1), GBT-A2 (G4BD-CNA2), GBT-B1 (G4BD-CNB1), GAD-A1 (G4AD-CNA1), GAD-A2 (G4AD-CNA2), and GAD-B1 (G4AD-CNB1) were constructed as follows. Plasmids containing inserts encoding 5 CN subunits CNA1 (SEQ ID NO:9; Cyert, et al., 1991), CNA2 (SEQ ID NO:11; Cyert, et al., 1991) and CNB1 (SEQ ID NO:13; Cyert, et al., 1992) were subjected to site-directed mutagenesis (Kunkle) to introduce a BamHI site just upstream of each subunit's initiation codon in the second reading frame. DNA prepared from the mutated plasmids was digested with BamHI and XhoI, and the resulting BamHI-XhoI fragments, each containing a full-length coding 10 sequence, were cloned into GBT or GAD that had been cut with BamHI and Sal 1. The resulting plasmids encoded in-frame fusions of the CN subunits with G4BD or G4AD.

Plasmids encoding CNA protein variants with truncated C-termini (GBT-A1\DeltaC, GBT-A2\(C \) were constructed by introducing stop codons after amino acid residues 509 (CNA1) and 502 (CNA2). The 44-residue deletion in GBT-A1\(Delta\)C removed the autoinhibitory domain of 15 CNA1, while the 102-residue deletion in GBT-A2ΔC removed both the autoinhibitory and the calmodulin-binding domains of CNA2 (Cyert, et al., 1991).

Plasmid GBT-FKBP, containing an FK506 binding protein (FKBP) gene fused to the GAL4 binding domain, was constructed by introducing a BgIII site upstream of the initiation codon and a BamHI site downstream of the stop codon of FKBP12 (Heitman, et al.) and ligating the BgIII-BamHI fragment into GBT cut with BamHI.

Plasmid B1/YEp352 was constructed to contain the full coding sequence of CNB1 (SEQ ID NO:13, Cyert, et al., 1992) as a 1.4 kb BamHI-EcoRI fragment encompassing the sequence presented as SEQ ID NO:13 (812 bp; contains the coding sequence), in the multicopy plasmid YEp352(HIS), which is derived from YEp352 (URA) (Hill, et al.).

Plasmid CNI/YEp352(HIS) (also referred to as CNIH) was constructed by ligating a 3.16 kb MunI-EcoRV fragment, containing the full coding sequence of CNI, from plasmid CN17.1 (construction described below) into YEp352(HIS) cut with EcoRI and Smal. Plasmids CNI/YEp352(TRP) (also referred to as CNIT) and CNI/YEp352(URA) (also referred to as CNIU) were similarly constructed using the 3.16 kb fragment and YEp352(TRP) or 30 YEp352(URA), respectively (Hill, et al.).

Plasmids A1/YEp351 and A2/YEp352 were constructed to contain the full coding sequences of CNA1 (SEQ ID NO:9, Cyert, et al., 1991) in YEp351 (Hill, et al.) and CNA2 (SEQ ID NO:11, Cyert, et al., 1991) in YEp351 and YEp352, respectively. A1/YEp351 was constructed by ligating a 2.9 kb SacI-HindIII fragment from clone CNA1 (Cyert, et al., 1991)

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into YEp351(HIS) cut with Sacl and HindIII. A2/YEp352 was similarly constructed by ligating a 3 kb Spel-HindIII fragment from clone CNA2 (Cyert, et al., 1991) into YEp352(HIS) cut with Xbal and HindIII.

All GAL4-CN fusions were verified by DNA sequencing (Sanger, et al.) using "SEQUENASE 2.0" sequencing kits (United States Biochemical, Cleveland, OH), and were subjected to the following tests. The functionality of the fusion proteins was assayed by determining whether they could complement the appropriate cn mutant phenotypes, using assays to measure the sensitivity to pheromone and Mn²⁺ (Reneke, et al., Cyert, et al., 1991). All of the GAL4-CN fusions were functional in this assay.

The fusion proteins were also tested for their ability to activate the reporter gene in the absence of the complementary GAL-4 domain fusion (i.e., in the presence of the complementary GAL4 domain not fused to a second protein, for example, G4BD-A1 vs G4AD) using the two-hybrid interaction assay described below. Only GBT-B1 and GAD-A1 were able to activate the reporter gene at low levels without the complimentary GAL-4 domain fusion – assays with the other fusion proteins in the absence of the complimentary GAL-4 domain fusion showed no detectable levels of expression.

The two-hybrid interaction assay was also used to test the ability of the fusions to interact specifically with another fusion containing complimentary GAL4 and CN domains (e.g., G4BD-A1 interacting with G4AD-B1). All CN hybrids were able to react specifically and result in an activation of the reporter gene that was clearly detectable above background. The high specificity witnessed in these experiments indicates that the GAL4 two-hybrid system can reliably be used to assay interactions between CN and other proteins.

D. Yeast GAL4 Two-Hybrid System for Detecting Protein-Protein Interaction

In the library screen, described in more detail in Example 1A, the yeast strain Y190, harboring the hybrid plasmid carrying the GAL4 binding domain fused to the A1 subunit of calcineurin (G4BD-CNA1), was transformed with fusion libraries carrying yeast genomic DNA Sau3AI fragments fused to the GAL4 activation domain. Transformants that were able to express the reporter genes, i.e., able to grow on -His + 3-AT and to score blue in β -gal assay, were selected as candidate positives. These candidate positives potentially contain library DNA fragments encoding proteins that physically interact with CNA1.

In another application described herein, the two-hybrid system was used to test for interactions between CNA (fused to one of the GAL4 domains) and CNB1 (fused to the other GAL4 domain), and between CNA and FKBP. Additional experiments tested a clone, CN1c,

isolated using the library screen, against a series of proteins fused to the complementary GALA domain under various conditions to test whether CN1c interacts with CNA subunits, and if so, how the interactions are affected by various conditions.

5 E. Color Development (β -gal) Assay

Yeast reporters harboring both G4BD and G4AD fusions (and a third non-fusion plasmid in some cases) were monitored for β -gal activity as follows. Purified yeast transformants were patched onto selective plates with or without other test reagents. After growing 3 days at 30°C, colonies were lifted onto nitrocellulose filters, permeabilized in liquid nitrogen as above, placed on Whatman No. 1 paper in petri dishes containing 0.1% X-Gal in Z buffer (see above), and incubated at 30°C for 12 hours. Blue color begins to appear in positive colonies between about one half and ten hours into the incubation period.

Exemplary images obtained using the color development assay are presented in Figures 2A, 3A, 4A, 4C, 5A, 5C, 6A, 6C, 6F, 7A, 7C, 7E, 7G, 8A, 9A and 9C.

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F. Growth Assay

A growth assay, applicable to yeast strains Y190 and Y153b1 which carry both GAL-HIS3 and GAL-lacZ reporters, was sometimes used as a complement to the color assay described above. Yeast transformants were streaked onto selective plates containing 40-50 mM 3-AT and no Histidine, and incubated at 30°C for 3-7 days. Growth (corresponding to the level of HIS3 expression) was monitored as an indicator of the interaction between fusion proteins. In cases where both assays were used, the amount of cell growth typically correlated well with the color intensity in the β -gal assay.

25 G. Yeast Growth, Drug Treatment

Yeast were typically grown in YPD (rich non-selective) or synthetic complete (SC) medium with selected component drop-outs, depending on the plasmid introduced, following standard procedures (Sherman, et al., Ausubel, et al.).

Experiments utilizing treatment with drugs or additives were performed by including the drug or additive in the medium. For plating, the agar was autoclaved, allowed to cool to 50° C, and the drug or additive was added before pouring the plates. Unless otherwise indicated, drugs and additives were added to result in the following final medium concentrations: FK506: 1 μ g/ml, cyclosporin A: 10 μ g/ml, rapamycin: 10 ng/ml, and hygromycin B: 40μ g/ml.

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H. Antibodies

Polyclonal and monoclonal antibodies, for use in the present invention, can be prepared by standard methods (Harlow, et al.) utilizing the CNI polypeptides of the present invention, for example, a substantially purified CNI/β -galactosidase fusion protein (Example 9). Antibodies can also be generated by recombinant techniques (Cabilly, et al.; Better, et al.; Skerra, et al.). In addition to whole antibody molecules, antibody fragments retaining the immunological specificity of the whole antibody may also be used in the practice of the present invention (e.g., Fab and F(ab')₂ fragments of IgG (Pierce Chemical, Rockford, IL)). The antibodies can be purified by standard methods to provide antibody preparations which are substantially free of serum proteins that may affect reactivity (e.g., affinity purification (Harlow et al.)).

EXAMPLE 1

Isolation of CNIc

A. Library Screening

Yeast strain Y190 was transformed with pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid using the transformation protocol described by Schiestl, et al. Transformants were selected, colony purified, and a single transformant was selected to make (Y190 GBT-A1)-competent cells, following the procedure described in Guthrie, et al..

The three pGAD yeast fusion libraries described above, carrying fusions between G4AD and yeast genomic DNA Sau3Al fragments in each reading frame, were then used to transform (Schiestl, et al.) the Y190 GBT-A1-containing cells. Transformants were plated onto SC-Trp-Leu-His plates containing 40 mM 3-aminotriazole (3-AT; Sigma Chemical Co., St. Louis, MO) and incubated at 30°C for 6 days to screen for HIS+ colonies (Durfee, et al.).

His $^{+}$ colonies were replica plated onto nitrocellulose filters (Schleicher & Schuell, Keene, NH), frozen in liquid nitrogen for approximately 30 seconds, and incubated at 30°C for 12 hours with Z buffer (see above) containing the chromogenic substrate X-Gal (0.1%) to assay β -gal activity (Breeden, et al.).

Candidate positive (blue) colonies were re-streaked for single colonies. Single colonies were purified and retested using the above protocol. Colonies which reproducibly tested positive were screened using PCR with primers directed against the internal portion of GAL-4 (i.e. the portion between the DNA binding domain and the activation domain). The sequences of the primers, G4-PCR-A and G4-PCR-B, are given as SEQ ID NO:22 and SEQ ID NO:23,

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respectively. Colonies yielding a PCR product were identified as containing intact GAL4, and were eliminated.

The GBT-A1 TRP* plasmid was eliminated by growing in Trp* liquid media for 2-3 days, plating on -Leu media and then replica-plating on -Leu and -Trp plates to identify and eliminate colonies that had lost the GBT-A1 plasmid, yet still gave a positive signal.

Plasmid DNA was extracted from the remaining Leu⁺ candidates. The plasmid DNA was transformed into E coli JBe181 and plated on -Leu media to select for library plasmids. The library plasmids isolated by this method were introduced back to the yeast reporter strains either alone or with test G4BD fusions: GBT, GBT-A1, and AS-lamin.

A parallel specificity assay was conducted by mating. Candidate strains, as described above, were 3-AT growth positive and X-gal positive when both the library and GBT-A1 plasmids were present. After elimination of the GBT-A1 plasmid from these strains, strains that were Leu* Trp* 3-AT growth and β -gal were mated to the following strains: Y187 (MATa) carrying GBT, GBT-A1, or AS-lamin, and the diploids were assayed.

Among the 3-AT positive, β -gal positive candidates identified by the secondary screening method just described, one clone (III-21S, later termed GAD-CNIc) was specifically positive in conjunction with GBT-A1 in both the transformation assay and the mating assay.

B. <u>Sequence of CNIc</u>

Clone III-21S was sequenced as above. The sequence is presented herein as SEQ ID NO:1, and a schematic representation of the clone is shown in Figure 1A. The Sau3AI library insert encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GAL4, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product.

25 Accordingly, the clone was termed CNIc, with the lowercase "c" representing "c-terminal".

The stippled portion between GAL-4AD and CNIc in Figure 1A represents the linker discussed in Materials and Methods, above. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons, and Sau3AI restriction sites.

30 C. Isolation of a Full Length Clone

A ³²P-labeled CNIc probe was generated from the 1.22 kb CNIc insert of clone III-21S by polymerase chain reaction (PCR) using primers represented as SEQ ID NO:7 and SEQ ID NO:8. The probe was used to map the gene to the right arm of chromosome 11 by hybridization screening (Sambrook, *et al.*) a panel of λ clones (American Type Culture

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Collection (ATCC), Rockville, MD) spanning the entire yeast genome. Two clones, 70500 and 70590, gave positive hybridization signals. A phage lysate of clone 70500 in λ MG3 was obtained from the ATCC, was amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments.

The phage DNA was digested with SacI, yielding a 7.1 kb fragment containing the entire CNI gene. This fragment was cloned into "BLUESCRIPT SK-" (Stratagene, La Jolla, CA) cut with SacI, yielding plasmid CNI7.1. Plasmid CNI7.1 was digested with Munl and EcoRV, releasing a 3.16 kb fragment containing the entire coding sequence of CNI. The 3.16 kb fragment was then cloned into each of YEp352(HIS), YEp352(TRP), YEp352(URA), and "BLUESCRIPT SK-", each cut with EcoRl and Smal, yielding plasmids CNIH, CNIT, CNIU and CNI3.2, respectively. The sequence of the 3.16 kb Munl/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4. The Munl site is at nucleotide 100 of SEQ ID NO:4, and the EcoRV site is at nucleotide 3263 of SEQ ID NO:4.

A schematic diagram of the sequence presented as SEQ ID NO:4 is shown in Figure 1B. This sequence contains the 3.16 kb MunI/EcoRV fragment used in many of the experiments described herein (depicted in Figure 1B as the portion between the MunI and EcoRV sites), which contains the entire 2.75 kb coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

Figure 13 also shows the location of certain features of the sequence. For example, "PEST" motifs (Rogers, et al., Dice) are indicated by bars over the corresponding sequence.

A search of known DNA and protein sequences turned up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin-binding protein.

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EXAMPLE 2

Binding of CNIc to Calcineurin

Y190 yeast carrying the plasmids indicated below were assayed for β -gal activity by color development assay described above to determine the specificity of binding of CNIc to subunits of calcineurin.

30 Exemplary data, in the form of images of filters having yeast colony replicas that had undergone the β-gal color development assay are shown in Figure 2A. The legend for Fig. 2A is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs. The constructs are as follows: 20:GBT-A1 and

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GAD-B1, 21:GBT-A1 and GAD-CNIc, 22:GBT-A1 and GAD-CNIc, 23:GBT-A1 and GAD-CNIc 24:GBT-A2 and GAD-CNIc 25:AS-lamin and GAD-CNIc 26:GBT and GAD-CNIc.

Yeast colonies used in the assay were derived by several different methods. Those at location 22 were purified colonies from the original library screen, those at 21 were colonies transformed with mini-prep DNA of the isolated GAD-CNIc plasmid, and the remaining colonies (23, 24, 25 and 26) were transformed with maxi-prep (Qiagen, Chatsworth, CA) DNA of GAD-CNIc.

A comparison of the intensities of the blue β -gal reaction product indicates that CNIc interacted strongly with CNA1 regardless of the source of the CNIc plasmid DNA (20, 21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with GAD-CNIc (26) showed a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact were used as a positive control for the assay (20).

In summary, the data above show that CNIc interacted specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments was conducted using constitutively-active CNA subunits, as well as calcineurin subunit CNB1. As described in Materials and Methods, above, CNA1ΔC and CNA2ΔC were each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A.

The legend for Fig. 3A is shown in Figure 3B. Locations of yeast colonies expressing specific constructs are as follows: 27:GBT and GAD-CNIc, 28:GBT-A1ΔC and GAD-CNIc, 29:GBT-A2ΔC and GAD-CNIc, 30:GBT-A1ΔC and GAD, 31:GBT-A2ΔC and GAD, 32:GBT-B1 and GAD-CNIc, 33:GBT-B1 and GAD, 34:GBT-A1ΔC and GAD-B1, and 35:GBT-A2ΔC and GAD-B1.

The data show that GBT-A1 Δ C and GAD-CNIc (28) gave a definite positive signal, while GBT-A2 Δ C and GAD-CNIc (29) was weaker, though still detectable above its background (i.e. GBT-A2 Δ C and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) was not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 Δ C and GAD-B1 (34) and GBT-A2 Δ C and GAD-B1 (35) gave strong signals.

These data show that CNIc interacted specifically with CNA1 Δ C and CNA2 Δ C, but not with CNB1.

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EXAMPLE 3

Effects of Immunosuppressant Drugs on Binding of CNIc to Calcineurin in B1. B1 Deletion and B1 Overproducing Yeast Strains

Three yeast strains carrying the plasmids indicated below were assayed for β -gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments were performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data, in the form of filter images produced as above are shown in Figures 4A, 4C, 5A and 5C. Plates used to make the filters shown in Figs. 4A and 4C were replicas from one master plate, while plates used to make the filters shown in Figs. 5A and 5C were replicas from another plate. The plates used to generate filters shown in Figs. 4A and 5A were without FK506, while the plates used to generate filters shown in Figs. 4C and 5C contained 1 µg/ml FK506.

The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Since the imaged filters in Figs. 4A and 4C were from replica plates, corresponding locations on each filter contain material from the same yeast colonies. Accordingly, the locations are referred to by the same "base" numbers in the legends. To facilitate reference to a specific location on a specific filter, the base numbers are followed by a lowercase letter that is different for each of the individual filters. For example, in the present figure, "a" follows the base numbers to identify locations on the filter shown in Fig. 4A, while a "b" follows the base numbers to identify locations on the plate in Fig. 4C. This labeling scheme is used in other experiments detailed herein where multiple filter lifts are shown.

The interactions of various combinations of proteins expressed by constructs indicated below was studied in three yeast strains. Strain Y153b, at 36-40, is null for the CNB1 subunit of calcineurin. Strains Y190 (41-46) and Y526 (47-51) are wild-type for CNA1, CNA2 and CNB1.

Hybrid proteins expressed by colonies at specific locations are as follows: 36:GBT-A1 and GAD-CNIc, 37:GBT-A2 and GAD-CNIc, 38:GBT-A1ΔC and GAD-CNIc, 39:GBT and GAD-CNIc, 40:GBT-A2ΔC and GAD-CNIc, 41:GBT-A1 and GAD-CNIc, 42:GBT and GAD-CNIc, 43:GBT-A2 and GAD-CNIc, 44:GBT-A1ΔC and GAD-CNIc, 45:GBT-A1 and GAD-B1, 46:GBT-A2ΔC and GAD-CNIc, 47:GBT-A1 and GAD-CNIc, 48:GBT and GAD-CNIc,

49:GBT-A2 and GAD-CNIc, 50:GBT-A1 Δ C and GAD-CNIc, and 51:GBT-A2 Δ C and GAD-CNIc.

Yeast strain Y526 was used for all experiments shown in Figs. 5A, 5B, 5C and 5D.

The expression vector B1/YEp352(HIS) was not used in strains Y190 or Y153b1 because they

are HIS⁺ in the absence of 3-AT.

The base numbers in Figures 5B and 5D correspond to locations of colonies expressing the following constructs: 52:GBT-A1, GAD-CNIc and YEp352, 53:GBT-A1, GAD-CNIc and B1/YEp352, 54:GBT-A2, GAD-CNIc and YEp352, 55:GBT-A2, GAD-CNIc and B1/YEp352, 56:GBT, GAD-CNIc and B1/YEp352, 57:GBT-A1\DeltaC, GAD-CNIc and B1/YEp352, and 58:GBT-A2\DeltaC, GAD-CNIc and B1/YEp352.

A comparison of data shown in Fig. 4A (no added drugs) shows the effect of deleting the endogenous host CNB1 gene on interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1ΔC and CNA2ΔC. Note that interactions in panels 36a-40a (CNB1 null strain) were all stronger (with the exception of the negative control in 39) than interactions in corresponding panels 41a-51a (strains wild-type for CNB1). This result indicates that interaction between CNIc and CNA subunits were enhanced by the deletion of the CNB1 subunit.

Comparison of corresponding panels in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA Δ C interactions. The drug enhanced interactions under all except control (39, 42 and 48) conditions. The effect was most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhanced CNIc-CNA/CNADC interactions under conditions where the CNB1 subunit was overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNADC interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) had reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhanced interactions in all colonies, except the negative controls (56).

Taken together, the above data demonstrate that the interaction of CNIc with CNA and CNA acc was markedly enhanced by FK506. The interaction was also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction was overcome by the stimulatory effect of FK506.

In light of the effects of FK506 on CNIc-CNA/CNA Δ C interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H. Filters shown in

Figs. 6A-6F were from replica plates, as were those in Figs. 7A-7H. Colonies shown in Figs. 6A and 7A were plated without drugs; those in Figs. 6C and 7C were plated with FK506 (1 μg/ml), those in Figs. 6E and 7E with CsA (10 μg/ml), and those in Fig. 7G with rapamycin (10 ng/ml). Yeast strains used were as follows: In Figs. 6A-6F, panels 59-63 were Y153b1, 64-69 were Y190, and 70-74 were Y526. In Figs. 7A-7H, panels 77-83 were Y526, and panels 75 and 76 were Y190.

The base numbers in Figures 6B, 6D and 6F correspond to locations of colonies expressing the following constructs: 59:GBT-A1 and GAD-CNIc, 60:GBT-A2 and GAD-CNIc, 61:GBT-A1\Delta C and GAD-CNIc, 62:GBT and GAD-CNIc, 63:GBT-A2\Delta C and GAD-CNIc, 64:GBT-A1 and GAD-CNIc, 65:GBT and GAD-CNIc, 66:GBT-A2 and GAD-CNIc, 67:GBT-A1\Delta C and GAD-CNIc, 68:GBT-A1 and GAD-B1, 69:GBT-A2\Delta C and GAD-CNIc, 70:GBT-A1 and GAD-CNIc, 71:GBT and GAD-CNIc, 72:GBT-A2 and GAD-CNIc, 73:GBT-A1\Delta C and GAD-CNIc, and 74:GBT-A2\Delta C and GAD-CNIc.

The base numbers in Figures 7B, 7D, 7F and 7H correspond to locations of colonies expressing the following constructs: 75:GBT-FKBP and GAD, 76:GBT-FKBP and GAD-CNIc, 77:GBT-A1, GAD-CNIc and YEp352, 78:GBT-A1, GAD-CNIc and B1/YEp352, 79:GBT-A2, GAD-CNIc and YEp352, 80:GBT-A2, GAD-CNIc and B1/YEp352, 81:GBT, GAD-CNIc and B1/YEp352, 82:GBT-A1ΔC, GAD-CNIc and B1/YEp352, and 83:GBT-A2ΔC, GAD-CNIc and B1/YEp352.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. The constructs and yeast strains at corresponding locations were the same. As expected, the β -gal signal was also essentially equivalent between the two sets. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A was also effective in enhancing interaction of CNIc with CNA and CNA Δ C. Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through inhibition of calcineurin activity (Cyert).

Similarly, data shown in Figures 7A and 7C are essentially equivalent to those in Figures 5A and 5C, except that a top panel has been added in Figs. 7A-H. As above, the corresponding panels show the same constructs and yeast strains. The added panels (75 and 76) assessed the interaction of an FK506 binding protein (FKBP) with CNIc, and indicate that there were no detectable interactions between these proteins. Results in Fig. 7E demonstrate that cyclosporin A had a similar effect to FK506 in cells overexpressing CNB1 — that is, it enhanced the interactions between CNIc and CNA/CNAAC.

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In contrast, data presented in Fig. 7G show that the immunosuppressant rapamycin, which is known not to target calcineurin, had no detectable effect on CNIc-CNA/CNA\(\Delta\) interactions (compare Fig. 7G with Fig. 7A).

Taken together, the above data show that like FK506, cyclosporin A (CsA), but not rapamycin, also enhanced the interaction of CNIc with CNA and CNAΔC. CNIc didn't interact with FKBP with or without FK506.

EXAMPLE 4

Effects of CNI on FKBP/FK506 binding to Calcineurin

Y526 cells, carrying the plasmids indicated below, were grown in -Trp-Leu-His liquid media with or without FK506 (1 μ g/ml) until OD₆₀₀ reached about 1.0. Approximately the same number of cells, calculated based on OD₆₀₀ and equivalent to 1 ml of an OD₆₀₀=1 suspension, was harvested from each culture, washed once with ddH₂O, centrifuged briefly, and the pellet was resuspended in 30 μ l ddH₂O and transferred onto a nitrocellulose filter. The filters were frozen in liquid nitrogen as described above, placed in a 8.5 cm petri dish containing a sheet of Whatman No. 1 paper (Whatman International LTD, Maidstone, UK) in 1.6 ml Z buffer containing 0.1% X-Gal, and incubated at 30°C for 8 hours.

Figure 8A presents exemplary data from studies to assess the effect of CNI overexpression on FK506-mediated FKBP interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Locations of yeast colonies expressing specific constructs: 84:GBT-FKBP, GAD-A2 and YEp352, 86:GBT-FKBP, GAD-A2 and YEp352, 86:GBT-FKBP, GAD-A2 and CNI/YEp352, and 87:GBT-FKBP, GAD-A2 and CNI/YEp352. The cells at 85 and 87 were exposed to FK506, while those at 84 and 86 were not.

Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 showed no detectable interaction (84). In the presence of FK506, however, the proteins interacted (85), presumably because FK506 formed a complex with FKBP, which then bound CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI had no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiated, or enhanced the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

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Experiments illustrated in Figures 9A and 9B demonstrate that CNI overproduction had little or no effect on the binding of CNA2 to CNB1, providing support for the specificity of the stimulatory effect that CNI overproduction had on the FK506-dependent binding of FKBP to calcineurin. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively.

Locations of yeast colonies expressing the following constructs: 88:GBT-A2, GAD-B1 and YEp352, 89:GBT-A2, GAD-B1 and YEp352, 90:GBT-A2, GAD-B1 and CNI/YEp352, and 91:GBT-A2, GAD-B1 and CNI/YEp352. The colonies at 89 and 91 were exposed to FK506, while colonies at 88 and 90 were not.

Taken together, the above data show that overexpression of the full-length CNI clone markedly enhanced the FK506-dependent interaction of FKBP with CNA, although it didn't affect the interaction between CNA and CNB1.

EXAMPLE 5

Co-Immunoprecipitation of CNIc and CNA

Yeast BJ2407 harboring AS-CNIc, which carries an influenza hemagglutinin (HA) epitope tag (Wilson, et al.), and GAD-A2 (lanes 1, 3) or A2/YEp352 (lanes 2, 5), and strain MCY300-1 (cna1 cna2; lane 4) were grown in selective media to OD_{e00}=0.8. The cells were harvested, lysed, and immunoprecipitated in the presence of 25 µg/ml FK506 with anti-HA monoclonal antibody (obtained from M. Kirschner, Harvard University, Boston, MA; Wilson, et al.), following protocols described in Harlow, et al. The cell extracts (lanes 3-5) and the immune complex (lanes 1, 2) were resolved by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; Laemmli) followed by western blot with a rabbit anti-CNA2 polyclonal antibody generated using standard methods (Harlow, et al.). Bound anti-CNA2 antibody was visualized with the "ECL" kit (Amersham, Arlington Heights, IL) using goat antirabbit antibody. Molecular weight markers are indicated on the right in kD.

The results demonstrate that CNIc was capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. This independent, biochemical assay confirmed the results described above obtained using the two hybrid protein interaction assay — that is, that CNIc physically interacted with and bound CNA subunits.

Cell extracts of BJ2407 harboring AS-CNIc, and GAD-A2 or A1/Yep351, and Y153b1 harboring AS-CNIc and GAD-A1 were subjected to SDS-PAGE followed by western blot with anti HA antibody. The results showed that CNIc was present at very low levels *in vivo*.

The results are consistent with the observations that a limited amount of CNA2 was precipitated by anti-HA antibody recognizing the CNIc fusions, and that CNI contains PEST-like motifs, a feature of proteins with a short half-life in vivo (Rogers, et al.).

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EXAMPLE 6

Northern Blot of CNIc

Norther blots (e.g., Sambrook, et al.) of yeast total RNA were hybridized with a CNIc probe. Exemplary data are shown in Figure 11. 20 µg yeast RNA from YPH499 (lane 1) and MCY300-1 (lane 2) was resolved in a formaldehyde-agarose gel, transferred onto "HYBOND N*" membrane (Amersham, Arlington Heights, IL), and hybridized with 5x10° cpm/ml probe of the CNIc insert (1.22kb). A single message of approximately 2.9 kb was detected in both strains at about the same level following an 18-hour exposure on XAR5 film (Eastman Kodak, Rochester, NY).

The data indicate that CNI was a physiologically expressed gene encoding a 2.9 kb message in yeast.

EXAMPLE 7

Chromosome Mapping of CNIc

A yeast chromosome blot obtained from the ATCC was hybridized with probe of the CNIc insert following the Southern hybridization procedure described in Sambrook, et al. A positive hybridization signal was obtained with two ATCC yeast genomic λ clones derived from chromosome 11. Clone 70500 had a relatively strong signal, while clone 70590 had a somewhat weaker one. A phage lysates of clone 70500 was ordered from the ATCC, amplified, purified, restriction-mapped, and used as a DNA source for cloning full length CNI (Example 1).

EXAMPLE 8

CNI null Mutants

1. Construction of cni Null Mutation

A 5', 1.8 kb Bg/II-HindIII and a 3', 0.9 kb Xbal-Bg/II fragment of CNI were ligated into pRS305(LEU2) (Sikorski, et al.). The resultant plasmid had a deletion of a 2 kb HindIII-Xbal fragment from the coding sequence of CNI. This cni::LEU2 mutant was introduced into the genomes of yeast haploid strains YPH499 (Sikorski, et al.), MCY300-1 (cnal cna2) and DD12 (cnbl) (Cyert, et al., 1991, Cyert, et al., 1992) as well as two diploid strains.

Leucine prototrophs were isolated at high frequency from all strains, and hybridization analysis confirmed that the *cni::LEU2* allele had replaced the CNI gene. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains (even cni cn double mutants) can survive.

CNI was deleted from three yeast strains: YPH499 (WT), MCY300-1 (cna1 cna2), and DD12 (cnb1), resulting in cni strains LHy499, LHy300 and LHy12, respectively. Cells representing four colonies of each *cni* knockout strain and two colonies of each parent strain were grown in liquid YPD (Sherman, *et al.*) to saturation. Same numbers of cells from each culture were then plated onto YPD+Hygromycin B (40 μ g/ml) and growth was monitored at 30°C.

CNI deletions in each strain rendered that strain more resistant to hygromycin B. The effect was particularly pronounced in both MCY300-1 and DD12, suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B. The data indicate that deletion of CNI results in higher resistance to hygromycin B.

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EXAMPLE 9

Isolation of CNI/B-Galactosidase Fusion Protein

A CNI coding sequence is cloned into the λ gt11 vector (Stratagene, La Jolla, CA). The coding frame is cloned in-frame to the β -galactosidase coding sequences present in λ gt11. Bacterial lysogens infected either with lambda phage gt11 or with gt11/CNI are incubated in 32°C until the culture reaches to an O.D. of 0.4. Then the culture is incubated in a 43°C water bath for 15 minutes to induce gt11 peptide synthesis, and further incubated at 37°C for 1 hour. Bacterial cells are pelleted and lysed in lysis buffer (10 mM Tris, pH 7.4, 2 % "TRITON X-100" and 1% aprotinin). Bacterial lysates are clarified by centrifugation (10K, for 10 minutes, Sorvall JA20 rotor) and the clarified lysates are incubated with Sepharose 4B beads conjugated with anti- β -galactosidase (Promega).

Binding and elution of β -galactosidase fusion proteins are performed according to the manufacturer's instruction. Typically binding of the proteins and washing of the column are done with lysis buffer. Bound proteins are eluted with 0.1 M carbonate/bicarbonate buffer, pH 10.

While the invention has been described with reference to specific methods and embodiments, it will be appreciated that various modifications and changes may be made without departing from the invention.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: The Board of Trustees of the Leland Stanford Junior University
- (ii) TITLE OF INVENTION: Calcineurin Interacting Protein Compositions and Methods
- (iii) NUMBER OF SEQUENCES: 23
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Dehlinger & Associates
 - (B) STREET: 350 Cambridge Avenue, Suite 250
 - (C) CITY: Palo Alto
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 94306
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 23-OCT-1995 (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/328,322
 - (B) FILING DATE: 24-OCT-1994
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Sholtz, Charles K.
 - (B) REGISTRATION NUMBER: P38,615
 - (C) REFERENCE/DOCKET NUMBER: 8600-0151.41
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (415) 324-0880
 - (B) TELEFAX: (415) 324-0960
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1222 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: Sau3AI fragment containing CNIc coding sequence
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..918

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

		AGT Ser										_			_	48
		CCC Pro														96
		TCG Ser 35													TCA Ser	144
		CCG Pro													GCA Ala	192
		TCT Ser													_	240
		GCC Ala														288
		CCT Pro														336
		GAG Glu 115														384
		GAT Asp														432
		ACC Thr														480
		TCT Ser														528
AGT Ser	CTA Leu	CAC His	GCA Ala 180	ACA Thr	TCA Ser	GTG Val	TTA Leu	CCT Pro 185	AGT Ser	ACA Thr	ATA Ile	AGA Arg	CAG Gln 190	AAC Asn	AAT ABN	576
		TTC Phe 195														624
GCC Ala	TTT Phe 210	CCC Pro	AAA Lys	AGC Ser	CAA Gln	TCA Ser 215	TTA Leu	AAT Aan	TTC Phe	AAT Asn	AAG Lys 220	AAA Lys	CTA Leu	CCA Pro	ATA Ile	672
CTT Leu 225	AAA Lys	ATT Ile	AAT ABN	GAT Asp	AAC Aen 230	GTC Val	ATA Ile	CAA Gln	TCA Ser	AAC Asn 235	AGC Ser	TAA Aen	AGT Ser	AAT Asn	AAC ABN 240	720
AGA Arg	GTT Val	GAT Asp	AAT Aan	CCA Pro 245	GAA Glu	GAT Asp	ACA Thr	GTG Val	GAT Asp 250	TCT Ser	TCA Ser	GTC Val	GAT Asp	ATT Ile 255	ACA Thr	768

			GAT Asp 260											Trp		816
			AAC Asn												GCT Ala	864
			CGT Arg												AAG Lye	912
AGA Arg 305		AAT	GTAC	ATT A	ATTT	rcati	rc To	CGAC	AGAJ	TTO	CTAC	CAT	TTT	ACTT:	rgt	968
GTC	CTGTC	TA	TCAA?	TAGTO	A T	TAAC	TAT	GG#	CATI	ATT	TAGT	ATA	CAA	LTATA	ACACO	A 1028
TCA	ATCT	ATA	CATCO	CATAT	A 27	CTTGI	CGT	AA A	CATA	ccc	TTTT	EAAT	'AG	TACAC	CGAT	TT 1088
KAAA	CAAA	AA (CATGA	ATTA	AC GT	CTCAC	TTAC	CA	ATGAC	CTT	ATTI	TTTAT	GG (CTTG	CTTTA	AG 1148
ATTI	TTC	CAA (GTCA	ATTTI	rt G	TTTI	TCT	ACC	CTTC	CAA	ссто	CATCI	CA I	ACCT:	CTTC	CC 1208
TTT	CAAC	CA (GATC													1222

- (2) INFORMATION FOR SEQ ID NO:2:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 306 amino acide
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Asp Gln Ser Ser Asn Val Phe Ala Ser Lys Gln Leu Val Ala Asn Ile
1 5 10 15

Tyr Lys Pro Asn Gln Ile Pro Arg Glu Leu Thr Ser Pro Gln Ala Leu 20 25 . 30

Pro Leu Ser Pro Ile Thr Ser Pro Ile Leu Asn Tyr Gln Pro Leu Ser 35 40 45

Asn Ser Pro Pro Pro Asp Phe Asp Phe Asp Leu Ala Lys Arg Gly Ala 50 55

Ala Asp Ser His Ala Ile Pro Val Asp Pro Pro Ser Tyr Phe Asp Val 65 70 75 80

Leu Lys Ala Asp Gly Ile Glu Leu Pro Tyr Tyr Asp Thr Ser Ser Ser 85 90 95

Lys Ile Pro Glu Leu Lys Leu Asn Lys Ser Arg Glu Thr Leu Ala Ser 100 105 110

Ile Glu Glu Asp Ser Phe Asn Gly Trp Ser Gln Ile Asp Asp Leu Ser 115 120 125

Asp Glu Asp Asp Asn Asp Gly Asp Ile Ala Ser Gly Phe Asn Phe Lys 130 140

Leu Ser Thr Ser Ala Pro Ser Glu Asn Val Asn Ser His Thr Pro Ile 145 150 155 160 WO 96/12806 PCT/US95/13580

41

Arg Lys

- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 918 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: cDNA to mRNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: CINc coding sequence
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GATCAAAGTA GCAATGTCTT CGCATCCAAA CAGCTGGTCG CAAACATTTA TAAGCCCAAT 60 CAGATTCCAA GAGAATTAAC TTCTCCTCAG GCGTTACCAT TATCGCCCAT CACCTCACCA 120 ATTCTCAATT ACCAACCATT ATCAAACTCC CCGCCTCCAG ATTTTGATTT TGATCTAGCT 180 AAGCGCGGCG CAGCCGATTC TCATGCTATT CCTGTGGATC CTCCATCATA TTTTGATGTA 240 TTANAGGCCG ATGGGATTGA ATTGCCATAC TACGATACAA GTTCATCTAA AATTCCTGAA 300 CTANAACTAA ACAAATCTAG AGAGACATTG GCCAGCATTG AGGAGGACTC ATTCAATGGT 360 TGGTCTCAAA TTGATGACTT ATCCGACGAA GATGACAATG ATGGCGATAT AGCATCTGGT 420 TTCAACTTCA AGCTGTCAAC CAGTGCTCCG AGTGAGAACG TTAATTCACA CACTCCTATT 480 TTGCAGTCTT TAAACATGAG TCTTGATGGG AGAAAAAAA ATCGTGCCAG TCTACACGCA 540

TTTCGCGAAA	AGAGAAAA					918
GCAGCCTACT	CGGTTAACGT	TGCTAGTGAA	AACCGTGTAC	TGGACGACTT	TAAGAAAGCA	900
CCAAGAATGT	CATCAGATTC	CAAATTTGAT	TGGGAGGTAA	GCAAGAACCA	TGTTGACCCA	840
AGAGTTGATA	ATCCAGAAGA	TACAGTGGAT	TCTTCAGTCG	ATATTACAGC	ATTTTATGAT	780
AAACTACCAA	TACTTAAAAT	TAATGATAAC	GTCATACAAT	CAAACAGCAA	TAGTAATAAC	720
ATGCTAGGCA	GTAGTGACGA	AGATGCCTTT	CCCAAAAGCC	AATCATTAAA	TTTCAATAAG	660
ACATCAGTGT	TACCTAGTAC	AATAAGACAG	AACAATCAGC	ATTTCAATGA	CATAAACCAG	600

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3500 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double

 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNI coding sequence
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 376..3120
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CCGAACACTT CCTTCGAGAG AGTGCATTTT ACTATGTGAA CCAATTTTCC CTCTTTTCG	60
GTTTGCAAGT TCACCTGAAA AACTGCTTAA CACTACTAGC AATTGCCCTA TTGTCGTACG	120
AGGACTTTGC CAAATGTATT CCCGGCTGTT TGTAGTATAT ATACGCAGAT ATATAATAGC	180
GCCGTCTTTT TACCTCTTTG AGCGAATTGC CAAATATTGA CTCTTTTGTC TTATTTCGCT	240
ATCCCCATCT TATCAAAAAT GGGAACAACT CGTTGAAATA AGAGACAAGC AACAAGAAAG	300
ACAACCAACA GAAAGTTCCA TTCCGCACAA ATACGCTGGA ATCCCATAGA ATATTGCTTG	360
TTCCTCTATG ACTAC ATG CTC CAA TTC AAT ACA GAA AAT GAT ACT GTA GCT Met Leu Gln Phe Asn Thr Glu Asn Asp Thr Val Ala 1 5 10	411
CCA GTG TTT CCC ATG GAG CAA GAT ATA AAT GCA GCA CCT GAT GCC GTC Pro Val Phe Pro Met Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val 15 20 25	459
CCA CTG GTG CAG ACA ACA ACA CTA CAA GTC TTT GTA AAG CTT GCC GAA Pro Leu Val Gln Thr Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu 30 35	507
30 33	

	ATC Ile								603
	AAA Lys 80								651
	GAA Glu								699
	GTC Val								747
_	TTC Phe								795
	ATG Met								843
	ATC 11e 160								891
	GAT Asp								939
	CCA Pro	_							987
	CAA Gln								1035
	AAT Asn								1083
	TTC Phe 240					_	_		1131
	GA G Glu								1179
	TT T Phe								1227
	TCT Ser								1275
	CT G Leu								1323
	ACT Thr 320								1371

			GAA Glu						1419
			GTG Val 355						1467
			GAT Asp						1515
			ATT						1563
			CAC His						1611
			GAA Glu						1659
			TTG Leu 435						1707
			GAT Asp						1755
			TTT Phe						1803
_			AAA Lys						1851
			GGG Gly						1899
			ATT Ile 515						1947
			GAG Glu						1995
			GCA Ala						2043
			CCA Pro						2091
			ATG Met						2139
			GAA Glu 595						2187

ACC Thr 605	TCC Ser	AAG Lyb	AAG Lys	TAA naa	GAT Asp 610	CAA Gln	AGT Ser	AGC Ser	AAT Aen	GTC Val 615	TTC Phe	GCA Ala	TCC Ser	AAA Lys	CAG Gln 620	2235
CTG Leu	GTC Val	GCA Ala	AAC Aen	ATT 11e 625	TAT Tyr	AAG Lys	CCC Pro	TAA naA	CAG Gln 630	ATT Ile	CCA Pro	AGA Arg	GAA Glu	TTA Leu 635	ACT Thr	2283
TCT Ser	CCT Pro	CAG Gln	GCG Ala 640	TTA Leu	CCA Pro	TTA Leu	TCG Ser	CCC Pro 645	ATC Ile	ACC Thr	TCA Ser	CCA Pro	ATT Ile 650	CTC Leu	TAA	2331
TAC Tyr	CAA Gln	CCA Pro 655	TTA Leu	TCA Ser	AAC Asn	TCC Ser	CCG Pro 660	CCT Pro	CCA Pro	GAT Asp	TTT Phe	GAT Asp 665	TTT Phe	GAT Asp	CTA Leu	2379
GCT Ala	AAG Lys 670	CGC Arg	GGC Gly	GCA Ala	GCC Ala	GAT Asp 675	TCT Ser	CAT His	GCT Ala	ATT	CCT Pro 680	GTG Val	GAT Asp	CCT Pro	CCA Pro	2427
TCA Ser 685	TAT Tyr	TTT Phe	GAT Asp	GTA Val	TTA Leu 690	AAG Lys	GCC Ala	GAT Asp	GGG Gly	ATT 11e 695	GAA Glu	TTG Leu	CCA Pro	TAC Tyr	TAC Tyr 700	2475
GAT Asp	ACA Thr	AGT Ser	TCA Ser	TCT Ser 705	AAA Lys	ATT Ile	CCT Pro	GAA Glu	CTA Leu 710	AAA Lys	CTA Leu	AAC Asn	AAA Lyb	TCT Ser 715	AGA Arg	2523
GAG Glu	ACA Thr	TTG Leu	GCC Ala 720	AGC Ser	ATT Ile	GAG Glu	GAG Glu	GAC Asp 725	TCA Ser	TTC Phe	TAA Aan	GGT Gly	TGG Trp 730	TCT Ser	CAA Gln	2571
ATT	GAT Asp	GAC Asp 735	TTA Leu	TCC Ser	GAC Asp	GAA Glu	GAT Asp 740	GAC Asp	AAT ABN	GAT Asp	GGC Gly	GAT Asp 745	ATA Ile	GCA Ala	TCT Ser	2619
GGT Gly	TTC Phe 750	AAC Asn	TTC Phe	AAG Lye	CTG Leu	TCA Ser 755	ACC Thr	AGT Ser	GCT Ala	CCG Pro	AGT Ser 760	GAG Glu	AAC Asn	GTT Val	AAT Asn	2667
TCA Ser 765	CAC His	ACT Thr	CCT Pro	ATT	TTG Leu 770	CAG Gln	TCT Ser	TTA Leu	AAC ABn	ATG Met 775	AGT Ser	CTT Leu	GAT Asp	GGG Gly	AGA Arg 780	2715
AAA Lys	AAA Lys	AAT Asn	CGT Arç	GCC Ala 785	AGT Ser	CTA	CAC His	GCA Ala	ACA Thr 790	TCA Ser	GTG Val	TTA Leu	CCT Pro	AGT Ser 795	ACA Thr	2763
						CAT His										2811
AGT Ser	AGT Ser	GAC Asp 815	GAA Glu	GAT Asp	GCC Ala	TTT Phe	CCC Pro 820	AAA Lys	AGC Ser	CAA Gln	TCA Ser	TTA Leu 825	AAT Asn	TTC	AAT	2859
AAG Lys	AAA Lys 830	Leu	CCA Pro	ATA Ile	CTT Leu	AAA Lys 835	ATT	TAA neA	GAT Asp	AAC Asn	GTC Val 840	ATA Ile	CAA Gln	TCA Ser	AAC Asn	2907
AGC Ser 845	TAA naa	AGT Ser	AAT Asn	AAC Asn	AGA Arg 850	GTT Val	GAT Asp	TAA nea	CCA Pro	GAA Glu 855	GAT Asp	ACA Thr	GTG Val	GAT Asp	TCT Ser 860	2955

			ATT Ile												TCC Ser	3003
			TGG Trp 880												TAC Tyr	3051
			GTT Val												AAA Lys	3099
			GAA Glu				TAAG	TAC	TT ?	ATTTI	CATI	TC TC	CCGA	CAGAJ	A.	3150
TTG	CTACO	AT :	TTAC	TTTC	T GI	CCTC	TGAT	TCA	ATA	TOT	ACA	TAT!	ATT (GAC	ATTTTA	3210
TAGT	OATA	AA I	ATATA	CACC	CA TO)TAA:	TATA	CAI	CCAT	OTAT	ACTI	GTC	ATE	AAGA	TATCCC	3270
TTTT	raat?	AG :	PACAG	CGAT	AA TI	SAAA	ATA	CAT	GATT	OKA	GTTC	CAGT	CAC (CAATO	AGCTT	3330
ATTI	TATTA	GG (TTG	TTT	ra Da	TTTT	CCAA	GTC	CTAA	TTT	GTTT	TTT	TA I	ACGC1	TTGCAA	3390
ССТС	CATCI	CA A	ACCTI	CTTC	C TI	TGC	AGCA	GA1	CTT	CGAA	ACCA	TCT	GT 1	TAT	CTCTC	3450
AATO	CTG1	TC (CACI	TTCA	AT CA	TCGT	CTGG	GAA	AAGI	COAT	GGTA	AGGG	CG			3500

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 915 amino acids

 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val Pro Leu Val Gln

Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu Pro Ile Val Phe

Leu Lys Gly Phe Glu Thr Asn Gly Leu Ser Glu Ile Ala Pro Ser Ile

Leu Arg Gly Ser Leu Ile Val Arg Val Leu Lys Pro Asn Lys Leu Lys 65 75 80

Ser Ile Ser Ile Thr Phe Lys Gly Ile Ser Arg Thr Glu Trp Pro Glu

Gly Ile Pro Pro Lys Arg Glu Glu Phe Ser Asp Val Glu Thr Val Val 100

Asn His Thr Trp Pro Phe Tyr Gln Ala Asp Asp Gly Met Asn Ser Phe

Thr Leu Glu His His Ser Ser Asn Asn Ser Ser Asn Arg Pro Ser Met

145						120									Val			
Fro	Pro	Thi	: Al	la (31u 165	Pro	Pro	L	/B]	Asp	Aen 170	ser	Asn	Leu	Ser	Le 17	u A 5	вр
Ala	туг	Gli	1 A1 18	rg 1	Asn	Ser	Leu	S S	er :	Ser 185	Asp	Asn	Leu	Ser	Авл 190	Ly	s P	ro
Val	ser	Se:	r A1	вр '	Val	Ser	His	A 1	вр 2 00	Asp	Ser	Lys	Leu	Lev 205	Ala	11	e G	ln
Lys	Thr	Pr	o L	eu i	Pro	Ser	Ser 215	S	er .	Arg	Arg	Gly	Ser 220	Va)	Pro	A l	a ?	Asn
Phe 225	Hie		y A	e n	Ser	Leu 230	Sea	r P	ro	His	Thr	Phe 235	Ile	Set	c Asj	, Le	eu I	Phe 240
		Th	r P	he	Ser 245	Asn	Se	r G	ly	Ala	Thr 250	Pro	Ser	Pre	o Glu	1 G	ln (55	Glu
Aep	ABI	а Ту	r L	eu 60	Thr	Pro	Se	r L	ys.	Авр 265	Ser	Lys	Glu	Va	1 Pho 27	e I: O	le :	Phe
Arg	Pr	o G1 27	y A	ар	туг	Ile	• Ту	r 1	hr 80	Phe	Glu	Glr	Pro	11 28	e Se 5	r G	ln	Ser
туг	Pr 29	o G1	u S	er	Ile	Lys	3 Al 29	a #	Asn	Phe	Gly	, Sei	7 Va	G1	u Ty	r L	ув	Leu
30	5					311	,								r Il			
Gli	n Le	u P	ro :	lle	Lys 325	Va.	l Va	1 1	Arg	Lev) Pro	se:	r As	p Gl	y Se	r V	al 35	Glu
Gl	u Th	r G	lu i	Ala 340	Ile	a Al	a I)	e	Ser	Ly 8	в Авј	p Tr	p Ly	s As	p Le 35	u I iO	.eu	His
		3	55						300						en Al 55			
	31	70					3	15							hr Le			
38	5					39	U					•	_		ye A			
					40	2									ys L			
				420	,					7.	•				ro A 4			
		- 4	135						771	,					eu A 45			
S	er G 4	ly 1 50	Asp	Le	u Va	1 A	en L	ув 155	Авј	p Ph	e G	ln P	he G 4	lu V 60	al P	he	Val	Pro
S (Phe	Th	r Ae	n S	er 1 70	1e	Ar	g Le	eu H	is P.	ro A 75	sp 1	hr A	sn	Tyr	486
		le :	Lys	Al	a Hi 48	.e H	is T	rp	11	e Ly	/8 I 4	le C 90	y s L	eu /	Arg I	Leu	Se:	Ly:

Lye	Tyr	Gly	Авр 500	Asn	Arg	Lys	Hie	Phe 505	Glu	Ile	Ser	Ile	Asp 510	Ser	Pro
Ile	His	11e 515	Leu	Asn	Gln	Leu	Сув 520	Ser	His	Ala	Asn	Thr 525	Leu	Leu	Pro
ser	Tyr 530	Glu	ser	His	Ph€	Gln 535	Tyr	Сув	Asp	Glu	Авр 540	Gly	Asn	Phe	Ala
Pro 545	Ala	Ala	Asp	Gln	Gln 550	Asn	Tyr	Ala	Ser	His 555	His	Авр	Ser	Asn	11e 560
Phe	Phe	Pro	Lys	Glu 565	Val	Leu	Ser		Pro 570	Val	Leu	Ser	Pro	Asn 575	Val
Gln	Lув	Met	Aen 580	Ile	Arg	Ile	Pro	Ser 585	qaA	Leu	Pro	Val	Val 590	Arg	Aen
Arg	Ala	Glu 595	Ser	Val	Lys	Lув	Ser 600	Lув	Ser	Asp	Asn	Thr 605	Ser	Lys	Lys
Asn	Авр 610	Gln	Ser	Ser	Asn	Val 615	Phe	Ala	Ser	Lys	Gln 620	Leu	Val	Ala	Asn
11e 625	Tyr	Lye	Pro	Asn	Gln 630	Ile	Pro	Arg	Glu	Leu 635	Thr	Ser	Pro	Gln	Ala 640
Leu	Pro	Leu	Ser	Pro 645	Ile	Thr	Ser	Pro	11e 650	Leu	Asn	Tyr	Gln	Pro 655	Leu
Ser	Asn	Ser	Pro 660	Pro	Pro	Asp	Phe	Asp 665	Phe	Asp	Leu	Ala	Lys 670	Arg	Gly
Ala	Ala	Asp 675	Ser	His	Ala	Ile	Pro 680	Val	Asp	Pro	Pro	Ser 685	Tyr	Phe	Авр
Val	Leu 690	Lys	Ala	Двр	Gly	11e 695	Glu	Leu	Pro	Tyr	Tyr 700	Asp	Thr	Ser	Ser
Ser 705	Lys	Ile	Pro	Glu	Leu 710	Lys	Leu	Asn	Lys	Ser 715	Arg	Glu	Thr	Leu	Ala 720
Ser	lle	Glu	Glu	Авр 725	Ser	Phe	Asn	Gly	Trp 730	Ser	Gln	Ilė	Авр	Авр 735	Leu
Ser	Авр	Glu	Авр 740	Asp	Asn	Авр	Gly	Asp 745	Ile	Ala	Ser	Gly	Phe 750	Aen	Phe
Lys	Leu	Ser 755	Thr	Ser	Ala	Pro	Ser 760	Glu	Aen	Val	Aen	Ser 765	His	Thr	Pro
Ile	Leu 770	Gln	Ser	Leu	Asn	Met 775	Ser	Leu	Asp	Gly	Arg 780	Lys	Lув	Asn	Arg
Ala 785	Ser	Leu	Hie	Ala	Thr 790	Ser	Val	Leu	Pro	Ser 795	Thr	Ile	Arg	Gln	800
Asn	Gln	His	Phe	Asn 805	Авр	Ile	Aen	Gln	Met 810	Leu	Gly	Ser	Ser	Авр 815	Glu
Asp	Ala	Phe	Pro 820	Lув	Ser	Gln	Ser	Leu 825	Aen	Phe	Asn	Lув	Lув 830	Leu	Pro
Ile	Leu	Lys 835	Ile	Asn	Asp	Asn	Val 840	Ile	Gln	Ser	Asn	Ser 845	Asn	Ser	Asn

Asn Arg Val Asp Asn Pro Glu Asp Thr Val Asp Ser Ser Val Asp Ile 850 855

Thr Ala Phe Tyr Asp Pro Arg Met Ser Ser Asp Ser Lys Phe Asp Trp 865 870 875 880

Glu Val Ser Lys Asn His Val Asp Pro Ala Ala Tyr Ser Val Asn Val 885 890 895

Ala Ser Glu Asn Arg Val Leu Asp Asp Phe Lys Lys Ala Phe Arg Glu 900 905 910

Lys Arg Lys

- (2) INFORMATION FOR SEQ ID NO:6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2745 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: cDNA to mRNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: coding sequence of CNI
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATGCTCCAAT TCAATACAGA AAATGATACT GTAGCTCCAG TGTTTCCCAT GGAGCAAGAT 60 ATAAATGCAG CACCTGATGC CGTCCCACTG GTGCAGACAA CAACACTACA AGTCTTTGTA 120 AAGCTTGCCG AACCCATAGT GTTTTTAAAA GGATTTGAAA CTAACGGACT GTCTGAAATA 180 GCCCCCAGTA TCTTACGAGG ATCTCTTATC GTCAGGGTGT TGAAACCGAA TAAATTAAAA 240 AGTATATCGA TAACCTTCAA AGGAATATCC AGAACAGAGT GGCCGGAAGG TATACCACCG 300 AAGAGAGAAG AATTTCAGA TGTTGAAACT GTTGTCAATC ACACATGGCC ATTTTATCAG 360 420 GCGGATGACG GCATGAATTC TTTCACCTTA GAACATCACA GCTCAAATAA TTCGTCCAAT CGCCCATCTA TGAGCGATGA AGATTATCTA CTTGAAAAAA GCGGTGCTTC AGTATATATC 480 CCACCAACCG CTGAACCCCC TAAAGATAAT AGCAATCTAA GTCTGGATGC CTATGAGCGC 540 600 AACTCATTGT CATCCGATAA TTTGAGTAAC AAGCCAGTAT CAAGTGATGT TTCCCATGAC GACAGTAAAC TGTTGGCTAT TCAAAAGACA CCATTACCAT CATCTAGTCG AAGAGGATCG 660 GTACCGCCAA ATTTTCACGG TAACTCTTTG TCACCTCATA CCTTCATATC TGATTTGTTC 720 780 ACAAAAACAT TCAGTAATAG TGGCGCTACT CCAAGTCCTG AGCAAGAGGA TAACTATCTT ACACCATCCA AAGATTCTAA AGAAGTTTTT ATTTTTCGAC CGGGCGATTA TATTTACACT **B40** TTTGAACAGC CAATATCGCA ATCTTATCCA GAAAGTATAA AAGCCAATTT TGGTTCCGTG 900 960 GAGTATAAAC TGTCAATAGA CATAGAGAGG TTTGGCGCAT TCAAATCAAC TATACATACT

CAATTACCCA	TCAAAGTCGT	AAGGCTTCCT	TCTGATGGAT	CCGTAGAAGA	GACTGAAGCT	1020
ATTGCAATTT	CCAAGGACTG	GAAAGATCTT	CTTCATTATG	ACGTGGTAAT	TTTCTCGAAA	1080
GAGATCGTTT	TGAATGCATT	TTTACCCATC	GATTTCCATT	TCGCTCCTCT	AGATAAAGTT	1140
ACTCTGCATC	GTATTAGAAT	TTATCTAACA	GAGTCTATGG	AATACACTTG	TAATAGTAAT	1200
GGAAATCACG	AGAAGGCTCG	TAGATTAGAG	CCAACTAAAA	AGTTTCTGTT	GGCTGAACAT	1260
AACGGTCCTA	AACTGCCTCA	TATACCAGCT	GGTTCGAATC	CTTTGAAGGC	TAAAAATAGA	1320
GGGAACATCC	TCTTGGATGA	AAAATCCGGC	GATCTAGTTA	ACAAAGATTT	TCAGTTCGAG	1380
GTGTTTGTCC	CAAGCAAGTT	TACAAACAGT	ATACGGTTAC	ACCCTGATAC	AAATTATGAT	1440
AAAATCAAAG	CCCACCATTG	GATAAAAATT	TGCCTTCGTC	TTTCCAAGAA	GTACGGGGAC	1500
AATAGAAAAC	ATTTCGAAAT	AAGTATTGAT	TCTCCAATCC	AAATTTTATA	TCAACTATGC	1560
TCACACGCGA	ATACTTTGCT	ACCGAGCTAC	GAGAGTCATT	TCCAGTÄTTG	TGATGAAGAT	1620
GGTAATTTCG	CACCAGCAGC	AGATCAACAA	AATTACGCAA	GTCATCATGA	TTCCAATATT	1680
TTCTTCCCAA	AAGAAGTTCT	TTCGTCTCCC	GTTCTTTCAC	CTAACGTGCA	GAAGATGAAC	1740
ATTAGAATAC	CGTCTGATCT	TCCAGTAGTG	CGTAATAGAG	CTGAAAGCGT	AAAGAAAAGC	1800
AAGTCAGATA	ATACCTCCAA	GAAGAATGAT	CAAAGTAGCA	ATGTCTTCGC	ATCCAAACAG	1860
CTGGTCGCAA	ACATTTATAA	GCCCAATCAG	ATTCCAAGAG	AATTAACTTC	TCCTCAGGCG	1920
TTACCATTAT	CGCCCATCAC	CTCACCAATT	CTCAATTACC	AACCATTATC	AAACTCCCCG	1980
CCTCCAGATT	TTGATTTTGA	TCTAGCTAAG	CGCGGCGCAG	CCGATTCTCA	TGCTATTCCT	2040
GTGGATCCTC	CATCATATTT	TGATGTATTA	AAGGCCGATG	GGATTGAATT	GCCATACTAC	2100
GATACAAGTT	CATCTAAAAT	TCCTGAACTA	AAACTAAACA	AATCTAGAGA	GACATTGGCC	2160
AGCATTGAGG	AGGACTCATT	CAATGGTTCG	TCTCAAATTG	ATGACTTATC	CGACGAAGAT	2220
GACAATGATG	GCGATATAGC	ATCTGGTTTC	AACTTCAAGC	TGTCAACCAG	TGCTCCGAGT	2280
GAGAACGTTA	ATTCACACAC	TCCTATTTTG	CAGTCTTTAA	ACATGAGTCT	TGATGGGAGA	2340
AAAAAAATC	GTGCCAGTCT	ACACGCAACA	TCAGTGTTAC	CTAGTACAAT	AAGACAGAAC	2400
AATCAGCATT	TCAATGACAT	AAACCAGATG	CTAGGCAGTA	GTGACGAAGA	TGCCTTTCCC	2460
AAAAGCCAAT	CATTAAATTT	CANTANGANA	CTACCAATAC	AATTAAAATT	TGATAACGTC	2520
ATACAATCAA	ACAGCAATAG	TAATAACAGA	GTTGATAATC	CAGAAGATAC	AGTGGATTCT	2580
TCAGTCGATA	TTACAGCATT	TTATGATCCA	AGAATGTCAT	CAGATTCCAA	ATTTGATTGG	2640
GAGGTAAGCA	AGAACCATGT	TGACCCAGCA	GCCTACTCGG	TTAACGTTGC	TAGTGAAAAC	2700
CGTGTACTGG	ACGACTTTAA	GAAAGCATTT	CGCGAAAAGA	GAAAA		2745

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 30 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

PCT/US95/13580

WO 96/12806

	51	
	(D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: CNI-PRC-A	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:7:	
ССААААА	AG AGATCTCGGA TCAAAGTAGC	30
(2) INFO	RMATION FOR SEQ ID NO:8:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: DNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: CNI-PCR-B	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GGGTTTTT	CA GTGTCGACGA TTCATAGATC	30
(2) INFO	RMATION FOR SEQ ID NO:9:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 1964 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: unknown	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNAl coding sequence	
(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2861944	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GTTTGTTGCA TTTTGATATT CATCTATATC TATTTCAAAA TTTTTCATGT CATCGCCTCT 60

TGA	AACA	TGA	ATTT	TCCA	T TA	CTGA	AAAA	G AA	CGTA	CTAC	TGG	GAAA	CAA	AAGG	AAAAA	120
TGT	AATA	ATC	CTTT	aatg:	TT T	TTGA	ATCA	A GA	GGCA'	TAT	TAT	AAAA	GAA (CGAA	GCAAAG	180
CCT	'AATT	TAT	TTGC	ATTT:	TT A	AAGG:	TATT	A TT	CAAA	AAA	AGT'	rttt:	TTA (GATT	CTTTTT	240
TTT	TTGA	CGT	ATTA	GCTC	AG C	TGCC	ATAA	A AC	actc:	TCAA	CGC			CG Al		294
			TCT Ser													342
			Asp Asp													390
			ATT Ile													438
			AAG Lys 55													486
			AAT Asn													534
			AAG Lye				-									582
			AAA Lys													630
			GAT Asp													678
			GGT Gly 135													726
			AGA Arg													774
			TTG Leu													822
			AAG Lye													870
			TAC Tyr													918
			CCA Pro 215													966

			lle											AAT Aen		1014
														GAC Asp		1062
	Trp													GGT Gly		1110
GAA Glu	TTT Phe	GAT Asp	CAG Gln	AGC Ser 280	GAG Glu	GAT Asp	GAA Glu	TTC Phe	GTA Val 285	CCT Pro	AAC Aen	AGT Ser	TTG Leu	AGG Arg 290	GGT Gly	1158
TGC Cys	TCT	TTC Phe	GCC Ala 295	TTC Phe	ACT Thr	TTT Phe	AAA Lys	GCA Ala 300	TCA Ser	TGC Cys	AAG Lys	TTT Phe	TTG Leu 305	AAG Lys	GCA Ala	1206
														GCT Ala		1254
														TTA Leu		1302
ACC Thr 340	Met	TTC Phe	AGT	GCG Ala	CCA Pro 345	AAC Asn	TAC Tyr	CTG Leu	GAC Asp	ACA Thr 350	TAT Tyr	CAT His	TAA nea	AAA Lys	GCT Ala 355	1350
GCT Ala	GTG Val	TTA Leu	AAA Lyb	TAT Tyr 360	GAA Glu	GAA Glu	AAC Aan	GTC Val	ATG Met 365	AAC Asn	ATC Ile	AGG Arg	CAG Gln	TTT Phe 370	CAC His	1398
														TTC Phe		1446
														GTG Val		1494
														GAA Glu		1542
														AAG Lys		1590
														GAA Glu 450		1638
														GCT Ala		1686
														GTG Val		1734
														CTA Leu		1782

	GAA Glu										1830
	GAC Asp										1878
	GAG Glu 535										1926
 	 CCA Pro	 	TGAT	[AAA]	rc r 1	CATI	CATT	T T			1964

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 553 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Ser lys Asp Leu Asn Ser Ser Arg Ile Lys Ile Ile lys Pro Asn 1 10 15

Asp Ser Tyr Ile Lys Val Asp Arg Lys Asp Leu Thr Lys Tyr Glu 20 25 30

Leu Glu Asn Gly Lys Val Ile Ser Thr Lys Asp Arg Ser Tyr Ala Ser 35

Val Pro Ala Ile Thr Gly Lys Ile Pro Ser Asp Glu Glu Val Phe Asp 50 55 60

Ser Lys Thr Gly Leu Pro Ash His Ser Phe Leu Arg Glu His Phe Phe 65 70 75 80

His Glu Gly Arg Leu Ser Lys Glu Gln Ala Ile Lys Ile Leu Asn Met
85 90 95

Ser Thr Val Ala Leu Ser Lys Glu Pro Asn Leu Leu Lys Leu Lys Ala 100 105 110

Pro Ile Thr Ile Cys Gly Asp Ile His Gly Gln Tyr Tyr Asp Leu Leu 115 120 125

Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Glu Ils Asp Tyr Leu Phe 130 135

Leu Gly Asp Tyr Val Asp Arg Gly Ala Phe Ser Phe Glu Cys Leu Ile 145 150 155 160

Tyr Leu Tyr Ser Leu Lys Leu Ain Asn Leu Gly Arg Phe Trp Met Leu 165 170 175

Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phe Lys 180 185 190

Asn Glu Met Leu His Lys Tyr Asp Met Glu Val Tyr Asp Ala Cys Cys 195 200 205

Arg	Ser 210	Phe	Asn	Val	Leu	Pro 215	Leu	Ala	Ala	Leu	Met 220	Asn	Gly	Gln	Туг
Phe 225	Сув	Val	His	Gly	Gly 230	Ile	Ser	Pro	Glu	Leu 235	Lys	Ser	Val	Glu	Asp 240
Val	Asn	Lув	Ile	Авп 245	Arg	Phe	Arg	Glu	11e 250	Pro	Ser	Arg	Gly	Leu 255	Met
Сув	Двр	Leu	Leu 260	Trp	Ala	Авр	Pro	Val 265	Glu	Asn	Tyr	Авр	Asp 270	Ala	Arg
Asp	Gly	Ser 275	Glu	Phe	увр	Gln	<i>Ser</i> 280	Glu	Asp	Glu	Phe	Val 285	Pro	asa	Ser
Leu	Arg 290	Gly	Сув	Ser	Phe	Ala 295	Phe	Thr	Phe	Lys	Ala 300	Ser	Сув	Lув	Phe
Leu 305	Lys	Ala	Asn	Gly	Leu 310	Leu	Ser	Ile	Ile	Arg 315	Ala	His	Glu	Ala	Glr 320
Авр	Ala	Gly	Tyr	Arg 325	Met	Tyr	Lys	Asn	Asn 330	Lys	Val	Thr	Gly	Phe 335	Pro
Ser	Leu	Ile	Thr 340	Met	Phe	Ser	Ala	Pro 345	Asn	Tyr	Leu	Asp	Thr 350	Tyr	His
Asn	Lys	Ala 355	Ala	Val	Leu	Lys	Tyr 360	Glu	Glu	Asn	Val	Met 365	Aen	Ile	Arg
Gln	Phe 370	His	Met	5er	Pro	His 375	Pro	Tyr	Trp	Leu	Pro 380	Авр	Phe	Met	Asp
Val 385	Phe	Thr	Trp	Ser	Leu 390	Pro	Phe	Val	Gly	Glu 395	Lys	Val	Thr	Ser	Met 400
				405	Asn				410					415	
Ser	Glu	Pro	Lys 420	Ala	Ala	Glu	Glu	Thr 425	Val	Lys	Ala	Arg	Ala 430	Asn	Ala
Thr	Lys	Glu 435	Thr	Gly	Thr	Pro	Ser 440	Asp	Glu	Lys	Ala	Ser 445	Ser	Ala	Ιlε
	450				Arg	455					460				
465					Arg 470					475					480
				485	Lys				490					495	
Ala	Leu	Ala	Arg 500	Gly	Thr	Glu	Gly	Leu 505	Asn	G]u	Thr	Leu	Ser 510	Thr	Ph€
		515	•		Glu		520					525			
Leu	Ser 530	Glu	Val	Glu	Gln	Glu 535	Lys	Ile	Lys	Tyr	Tyr 540	Glu	Lys	Ile	Leu
Lys		Ala	Glu	Lys	Lys 550	Pro	Gln	Leu							

121	INFORMATION	EUB.	CEO	TD	NO.	11.
121	INFORMATION	rok	SEU	טג	NU:	11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2353 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNA2 coding sequence
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 262..2073
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

ATAGTCTATA AT	ACGTTTGA TACAG	CTAGA TATCGCTAGO	GCCAACATTG TCCC	CCTCTC 60
TTGATCAATG CT	TTTTTCG GCCCG	AGACA AATGAGAAA	TGTCCTAAAA ATAC	CTTTCA 120
TCAAGACTCC TA	TTTTTCCT TAGAA	AAAAC ATATATCCAA	CTGGAACAGT ATTAI	AGCCAA 180
TTGCTACGAT AC	AAACAAAA GGAGA	TATTC CTTCCCTCC	ATAGAGTCAC ACAGO	GAGCCA 240
GTACTTCTTC TT		Ser Ser Asp Ala	ATA AGA AAT ACT Ile Arg Asn Thr	
			AAA ACA GAG CGT Lys Thr Glu Arg 25	
Gln Ser Ser Ti			AGT ACA GTT GCT Ser Thr Val Ala 40	
			CTT ACA CAA TAT Leu Thr Gln Tyr 55	
CTA GAT GAC GO Leu Asp Asp G	GA AGA GTC GTA ly Arg Val Val 65	Ser Thr Asn Arg	AGA ATA ATG AAT Arg Ile Met Asn 70	AAA 483 Lys
			GAA GAG CTG TTC Glu Glu Leu Phe	
			GAT CAT TTC AAG ABP His Phe Lys 105	
Glu Gly Lys Lo	TG TCG GCT GCG eu Ser Ala Ala 10	CAG GCG GCC AGG Gln Ala Ala Arg 115	ATC GTT ACA CTT Ile Val Thr Leu 120	GCA 627 Ala
ACG GAA CTC To Thr Glu Leu Pi 125	TC AGC AAA GAA he Ser Lys Glu	CCC AAC CTT ATA Pro ABn Leu Ile 130	TCT GTT CCC GCC Ser Val Pro Ala 135	CCA 675 Pro

					GAT Asp										AAG Lyb	723
					GGA Gly 160											771
					AGA Arg											819
					CTG Leu											867
					AAG Lys											915
					TAC Tyr										GAA Glu	963
					CCC Pro 240											1011
					ATA Ile											1059
					TTC Phe											1107
					GAC Asp											1155
					GAA Glu											1203
					GCA Ala 320											1251
					TAT Tyr											1299
CTG Leu	CAA Gln	GAG Glu	ACT Thr 350	GGC Gly	CTG Leu	TTG Leu	TCC Ser	ATC Ile 355	ATC Ile	AGG Arg	GCA Ala	CAC His	GAG Glu 360	GCT Ala	CAA Gln	1347
GAC Asp	GCT Ala	GGT Gly 365	TAT Tyr	AGA Arg	ATG Met	TAC Tyr	AAA Lye 370	AAT Aen	ACC Thr	AAG Lys	ACT Thr	TTG Leu 375	GGC Gly	TTT Phe	CCC Pro	1395
TCT Ser	CTT Leu 380	TTG Leu	ACC Thr	CTT Leu	TTC Phe	AGT Ser 385	GCG Ala	CCT Pro	AAC Aan	TAC Tyr	TTG Leu 390	GAC Asp	ACC Thr	TAC Tyr	AAT Asn	1443
AAT Asn 395	AAG Lys	GCT Ala	GCC Ala	ATA Ile	TTG Leu 400	AAA Lys	TAC Tyr	GAA Glu	TAA nea	AAT Asn 405	GTT Val	ATG Met	TAA Asn	ATC Ile	AGA Arg 410	1491

											CCA Pro					1539
											AAA Lye					1587
											GAG Glu				Asp	1635
											AAA Lys 470					1683
											ACT Thr					1731
											AGG Arg					1779
											ATG Met					1827
											GAT Asp					1875
											AAG Lys 550					1923
											GAT Asp					1971
AAA Lys	TTA Leu	CCG Pro	CCT Pro	TCA Ser 575	CTA Leu	GAC Asp	GAA Glu	CTG Leu	AAA Lys 580	AAC Aen	GAA Glu	AAT ABN	AAG Lys	AAG Lys 585	TAC Tyr	2019
											GAT Asp					2067
AGC Ser	_	TAG	AGAAJ	AGC 2	CCT	TTT	CC AC	TGT)	CATA	A CTI	CAA91	DAAT	TAAC) KAT	STT	2123
GCAT	raat1	TA 7	CTAT	TTAC	A AC	CTAC	ATGO	TCC	CTCAF	ATG	CACA	GAAI	rca 1	TATAC	CGTTT	2183
TAT	ragg:	CT (TTC	TTAT	T T	AGTI	TTGI	TGI	ATCTO	TAT	GAAG	GTAT	TAT 1	TAT	ATGCAA	2243
AAA :	AAA1	TT T	LAA T?	TAT	T AT	GGAT	CCTI	C AC	raadi	TGT	ATA	ACGI	TT 1	TCAT	RAGGAG	2303
TGC	LAAT:	TAT	GAC	CCAC	C TI	CTA	TTG	GCZ	GAAC	CCG	TTCI	CAAD	TC			2353

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:

 (A) LENGTH: 604 amino acids

 (B) TYPE: amino acid

 (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:
- Met Ser Ser Amp Ala Ile Arg Amn Thr Glu Gln Ile Amn Ala Ala Ile
- Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro 20 25 30
- Ile Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr 35 40 45
- Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val
 50 60
- Val Ser Thr Asn Arg Arg Ile Met Asn Lys Val Pro Ala Ile Thr Ser 65 70 75 80
- His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg 85 90 95
- His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala 100 105 110
- Ala Gln Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys 115 120 125
- Glu Pro Asn Leu Ile Ser Val Pro Ala Pro Ile Thr Val Cys Gly Asp 130 135 140
- Ile His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly 145 150 155 160
- Asp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg 165 170 175
- Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu 180 185 190
- Asn Phe Asn Asp His Phe Trp Leu Leu Arg Gly Asn His Glu Cys Lys 195 200 205
- His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu His Lys Tyr 210 215 220
- Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro 225 230 235
- Leu Ala Ala Leu Met Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile 245 250 255
- Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe 260 265 270
- Arg Glu Ile Pro Ser His Gly Leu Met Cys Asp Leu Leu Trp Ala Asp 275 280 285
- Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu 290 295 300
- Asp Ile Val Asn Ser Lys Thr Met Val Pro His His Gly Lys Met Ala 305 310 315
- Pro Ser Arg Amp Met Phe Val Pro Amn Ser Val Arg Gly Cym Ser Tyr 325 330 335

- Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu 340 345 350
- Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met 355 360 365
- Tyr Lye Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe 370 380
- Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu 385 390 395 400
- Lys Tyr Glu Asn Asn Val Met Asn Ile Arg Gln Phe Asn Met Thr Pro 405 410 415
- His Prc Tyr Trp Leu Pro Asp Phe Met Asp Val Phe Thr Trp Ser Leu 420 425 430
- Pro Phe Val Gly Glu Lys Val Thr Glu Met Leu Val Ala Ile Leu Asn 435 440
- Ile Cys Thr Glu Asp Glu Leu Glu Asn Asp Thr Pro Val Ile Glu Glu 450 455 460
- Leu Val Gly Thr Asp Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala 465 470 480
- Thr Prc Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu 485 490 495
- Amp Amp Glu Him Arg Arg Lym Ala Leu Arg Amn Lym Ile Leu Ala Val 500 505 510
- Ala Lys Val Ser Arg Met Tyr Ser Val Leu Arg Glu Glu Thr Asn Lys 515 520 525
- Val Gln Phe Leu Lys Asp His Asn Ser Gly Val Leu Pro Arg Gly Ala 530 535 540
- Leu Ser Asn Gly Val Lys Gly Leu Asp Glu Ala Leu Ser Thr Phe Glu 545 550 555
- Arg Ala Arg Lys His Asp Leu Ile Asn Glu Lys Leu Pro Pro Ser Lev 565 570 575
- Asp Glu Leu Lys Asn Glu Asn Lys Lys Tyr Tyr Glu Lys Val Trp Gln 580 585 590
- Lys Val His Glu His Asp Ala Lys Asn Asp Ser Lys 595 600
- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 812 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full

CNB1 coding sequence

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 54..104

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 181..652

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
ACTTGGTAAC TCAATGGTGA TCAGAATCCA TAGAAGCATT TTTATTTCTT AAA ATG Met 1	56
GGT GCT GCT CCT TCC AAA ATT GTG GAT GGT CTT TTA GAA GAT ACA AAT Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr Asn 5 10 15	104
TGTATGTACA CTTCGGAGTG AGGAAAAGAA AGAAAGGGGA AATTAACCGA TTTTACTAAC	164
ACTGACACTT TGAACA GTT GAT AGA GAT GAA ATT GAA AGG TTA AGG AAG Val Asp Arg Asp Glu Ile Glu Arg Leu Arg Lys 1 5 10	213
AGA TTC ATG AAA TTA GAT AGA GAT AGC TCA GGG TCT ATT GAT AAA AAT Arg Phe Met Lys Leu Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn 15 20 25	261
GAA TTT ATG AGC ATT CCT GGC GTT TCG TCA AAC CCT CTT GCT GGA CGT Glu Phe Met Ser Ile Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg 30 35 40	309
ATA ATG GAG GTT TTC GAT GCT GAT AAT AGT GGG GAC GTG GAT TTT CAA 11e Met Glu Val Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln 45 50 55	357
GAG TTC ATC ACA GGA TTA TCC ATT TTC AGT GGG CGT GGG TCC AAG GAC Glu Phe Ile Thr Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp 60 65 70 75	405
GAA AAG TTA AGA TTC GCC TTC AAA ATC TAC GAC ATT GAC AAG GAC GGT Glu Lys Leu Arg Phe Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly 80 85 90	453
TTC ATA TCC AAT GGT GAG TTG TTC ATC GTG TTG AAG ATT ATG GTA GGT Phe Ile Ser Asn Gly Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly 95 100 105	501
TCT AAT CTG GAC GAT GAA CAG CTG CAA CAG ATA GTA GAT AGG ACG ATA Ser Asn Leu Asp Asp Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile 110 115 120	549
GTG GAA AAC GAT AGC GAC GGC GAC GGA CGT TTA AGT TTC GAG GAG TTT Val Glu Asn Asp Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe 125 130 135	597
AAG AAT GCT ATC GAA ACC ACA GAA GTG GCC AAG AGT CTG ACA TTG CAA Lys Asn Ala Ile Glu Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln 140 145 150	645
TAC GATGTGTAAG ACTAGGGGAC ACTTCATTCA TTTATGGTAT GCCAATATTT Tyr Asp	698

TTAAGAAAG AAGAATAATA CGCGATATTG TTTTTTAAGG AAGGAACGCA CACTCGCCCA 758
GTTAGAGTGC TGATGATATA TACATATATA TATGTATATG TAACAAACAA 7AAG 812

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr 1 5 10 15

Asn

- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 157 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Val Amp Arg Amp Glu Ile Glu Arg Leu Arg Lym Arg Phe Met Lym Leu
1 10 15

Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn Glu Phe Met Ser Ile 20 25 30

Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Het Glu Val Phe 35 40

Asp Ala Asp Asn Ser Cly Asp Val Asp Phe Gln Glu Phe Ile Thr Gly 50 55

Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp Glu Lys Leu Arg Phe 65 70 75 80

Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn Gly 85 90 95

Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly Ser Asn Leu Asp Asp 100 105 110

Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile Val Glu Asn Asp Ser 115 120 125

Amp Gly Amp Gly Arg Leu Ser Phe Glu Glu Phe Lym Amn Ala Ile Glu 130 140

Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp 145 150 155

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:

 - (A) LENGTH: 524 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: coding sequence of CNB1
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..524
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

															ACA Thr	48
			AGA Arg 20													96
			GAT Asp												AGC Ser	144
			GTT Val													192
			GAT Asp													240
			ATT Ile													288
			AAA Lys 100													336
			TTC Phe													384
			CTG Leu													432
AGC Ser 145	GAC Asp	GGC Gly	GAC Asp	GGA Gly	CGT Arg 150	TTA Leu	AGT Ser	TTC Phe	GAG Glu	GAG Glu 155	TTT Phe	AAG Lys	AAT Asn	GCT Ala	ATC Ile 160	480
			GAA Glu											GT		524

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 174 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr

Asn Phe Asp Arg Asp Glu lle Glu Arg Leu Arg Lys Arg Phe Met Lys 20 25 30

Leu Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn Glu Phe Met Ser 35

Ile Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Met Glu Val 50 60

Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln Glu Phe Ile Thr
65 70 75 80

Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp Glu Lys Leu Arg 85 90 95

Phe Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn 100 105 110

Gly Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly Ser Asn Leu Asp 115 120 125

Amp Glu Gln Leu Gln Gln Ile Val Amp Arg Thr Ile Val Glu Amn Amp 130 135 140

Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe Lys Asn Ala Ile 145 150 155 160

Glu Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp 165 170

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1812 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: DNA fragment containing CNAldeltaC coding sequence
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 286..1812

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GTTT	GTTGCA	TTTTGA	TATT C	ATCTA	TATC	TATTTC	aaaa	TTT	TTCA	TGT	CATC	GCCTCT	60
TGAAJ	ACATGA	ATTTTC	CAAT 1	CTGAA	AAAG :	AACGTA	CTAC	TGG	GAAA	CAA .	aagg	GAAAAA	120
TGTAT	TAAATC	CTTTAA	TGTT 1	TTGAA	TCAA	GAGGCA	TATT	TAT	AAAA	GAA	CGÅA	GCAAAG	180
CCTTT	TARTAT	TTGCTT	A TTAT	AAGGT.	ATTA '	TTCAAA	GAAA	AGT	TTTT	TTA (GATT	CTTTTT	240
TTTT	TGACGT	ATTAGC	TCAG C	TGCCA	TAAA A	aca ct c	TCAA	CGC			CG A Br L		294
	TTG AAT Leu Asn 5												342
	AAG GTT Lys Val			Lys									390
	AAA GTA Lys Val	Ile S											438
	ACA GGA Thr Gly				Авр С								486
	TTA CCT Leu Pro 70	Asn H											534
	CTT TCT Leu Ser 85												582
	TTG AGT Leu Ser												630
	TGT GGT Cys Gly	Asp I											678
	GTT GGC /al Gly				Glu II								726
	GTT GAT /al Asp 150			Phe S			-						774
Ser L	TTG AAG Leu Lys 165												822
	AG TGT												870
	CAC AAA His Lys		p Met										918

			CCA Pro 215													966
			ATC Ile													1014
			TTT Phe													1062
			GAT Asp													1110
_			CAG Gln													1158
			GCC Ala 295													1206
			TTA Leu	-										-		1254
			TAT Tyr													1302
			AGT Ser													1350
			AAA Lys													1398
			CAC His 375									Авр				1446
			CCT Pro													1494
			ATA Ile													1542
			GAG Glu													1590
			CCA Pro													1638
GAA Glu	ACC Thr	CGA Arg	AGA Arg 455	AAG Lys	GCT Ala	TTG Leu	AGA Arg	AAT Asn 460	AAG Lys	ATA Ile	TTA Leu	GCT Ala	ATT Ile 465	GCT Ala	AAA Lys	1686
GTT Val	TCA Ser	AGA Arg 470	ATG Met	TTT Phe	TCG Ser	GTG Val	CTA Leu 475	AGA Arg	GAA Glu	GAG Glu	AGC Ser	GAA Glu 480	AAA Lys	GTG Val	GAA Glu	1734

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TAT TTG AAA ACT ATG AAT GCC GGT GTC TTA CCT CGT GGT GCT CTA GCT 1782 Tyr Leu Lys Thr Met Asn Ala Gly Val Leu Pro Arg Gly Ala Leu Ala 490 CGT GGG ACT GAA GGT TTG AAT GAA ACG CTA 1812

Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu 505

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 509 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Het Ser Lys Asp Leu Asn Ser Ser Arg Ile Lys Ile Ile Lys Pro Asn

Asp Ser Tyr Ile Lys Val Asp Arg Lys Lys Asp Leu Thr Lys Tyr Glu 20 25 30

Leu Glu Asn Gly Lys Val Ile Ser Thr Lys Asp Arg Ser Tyr Ala Ser

Val Pro Ala Ile Thr Gly Lys Ile Pro Ser Asp Glu Glu Val Phe Asp

Ser Lys Thr Gly Leu Pro Asn His Ser Phe Leu Arg Glu His Phe Phe

His Glu Gly Arg Leu Ser Lys Glu Gln Ala Ile Lys Ile Leu Asn Met

Ser Thr Val Ala Leu Ser Lys Glu Pro Asn Leu Leu Lys Leu Lys Ala

Pro Ile Thr Ile Cys Gly Asp Ile His Gly Gln Tyr Tyr Asp Leu Leu

Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Glu Ile Asp Tyr Leu Phe

Leu Gly Asp Tyr Val Asp Arg Gly Ala Phe Ser Phe Glu Cys Leu Ile

Tyr Leu Tyr Ser Leu Lys Leu Asn Asn Leu Gly Arg Phe Trp Met Lau

Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phe Lys

Asn Glu Met Leu His Lys Tyr Asp Met Glu Val Tyr Asp Ala Cys Cys

Arg Ser Phe Asn Val Leu Pro Leu Ala Ala Leu Met Asn Gly Gln Tyr 210 215

Phe Cys Val His Gly Gly Ile Ser Pro Glu Leu Lys Ser Val Glu Asp

Val Asn Lys Ile Asn Arg Phe Arg Glu Ile Pro Ser Arg Gly Leu Met 250

- Cys Asp Leu Leu Trp Ala Asp Pro Val Glu Asn Tyr Asp Asp Ala Arg 260 265 270
- Amp Gly Ser Glu Phe Amp Gln Ser Glu Amp Glu Phe Val Pro Amn Ser 275 280 285
- Leu Arg Gly Cys Ser Phe Ala Phe Thr Phe Lys Ala Ser Cys Lys Phe 290 295 300
- Leu Lys Ala Asn Gly Leu Leu Ser 11e Ile Arg Ala His Glu Als Gln 305 310 315 320
- Asp Ala Gly Tyr Arg Met Tyr Lys &sn Asn Lys Val Thr Gly Phe Pro 325 330 335
- Ser Leu Ile Thr Met Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr His 340 345 350
- Asn Lys Ala Ala Val Leu Lys Tyr Glu Glu Asn Val Met Asn Ile Arg 355 360 365
- Gln Phe His Met Ser Pro His Pro Tyr Trp Leu Pro Asp Phe Met Asp 370 375 380
- Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Ser Met 385 390 395 400
- Leu Val Ser Ile Leu Aan Ile Cys Ser Glu Gln Glu Leu Aap Pro Glu 405 415
- Ser Glu Pro Lys Ala Ala Glu Glu Thr Val Lys Ala Arg Ala Asn Ala 420 425 430
- Thr Lys Glu Thr Gly Thr Pro Ser Asp Glu Lys Ala Ser Ser Ala Ile 435 440 445
- Leu Glu Asp Glu Thr Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala 450 455 460
- Ile Ala Lys Val Ser Arg Met Phe Ser Val Leu Arg Glu Glu Ser Glu 465 470 475 480
- Lys Val Glu Tyr Leu Lys Thr Met Asn Ala Gly Val Leu Pro Arg Gly 485 490 495
- Ala Leu Ala Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu 500 505
- (2) INFORMATION FOR SEQ ID NO: 20:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1767 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: DNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: DNA fragment containing CNA2deltaC coding sequence
 - (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 262..1767

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

ATA	GTCT	ATA	ATAC	GTTT	GA T	ACAG	CTAG	A TA	TCGC	TAGC	GCC	AACA'	TTG	TCCC	сстстс		60
TTG	ATCA	ATG	CTTT	TTTT	CG G	CCCG.	AGAC	AA A	TGAG	AAAA	TGT	CCTA	AAA .	ATAC	CTTTCA		120
TCA	AGAC	TCC	TATT	TTTC	CT T	AGAA.	AAAA	C AT	ATAT	CCAA	CTG	GAAC	AGT .	ATTA	AGCCAA		180
TTGCTACGAT ACAAACAAAA GGAGATATTC CTTCCCTCCC ATAGAGTCAC ACAGGAGCCA												:	240				
GTA	CTTC	TTC	TTGA	ACCC	GC A					GCT Ala 5						;	291
			GCC Ala													;	339
_			ACA Thr 30													;	387
			ACG Thr													•	435
			GGA Gly													•	483
			ATC 11e													į	531
			ATA													!	579
			TTG Leu 110													•	627
			TTC Phe													•	675
			TGC Cys														723
			GTT Val													•	771
			GTC Val													8	819
			TTG Leu 190													8	B 67

															AAT Asn	915
			CAC His												GAA Glu	963
			AAC Asn													1011
			GGT Gly			-										1059
			ААТ Авл 270													1107
			TGG Trp													1155
			ACT Thr													1203
			AAG Lys													1251
			TGT Cys													1299
_		_	ACT Thr 350								_					1347
			TAT Tyr													1395
			ACC Thr													1443
			GCC Ala													1491
			ATG Het		-			_								1539
			TGG Trp 430													1587
CTT Leu	GTC Val	GCA Ala 445	ATT Ile	CTA Leu	AAC ABN	ATC Ile	TGT Cys 450	ACT Thr	GAA Glu	GAT Asp	GAG Glu	CTG Leu 455	GAA Glu	AAC Asn	GAC Aвр	1635
ACC Thr	CCC Pro 460	GTC Val	ATT Ile	GAA Glu	GAA Glu	TTA Leu 465	GTT Val	GGT Gly	ACC Thr	GAT Asp	AAA Lys 470	AAA Lys	TTG Leu	CCA Pro	CAA Gln	1683

GCT GGT AAG TCG GAA GCA ACT CCA CAA CCA GCC ACT TCG GCG TCG CCT
Ala Gly Lys Ser Glu Ala Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro
475

480

485

490

1767

AAA CAT GCT TCC ATT TTA GAT GAC GAA CAT CGA AGG Lys His Ala Ser Ile Leu Asp Asp Glu His Arg Arg 495 500

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 502 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Met Ser Ser Amp Ala Ile Arg Amn Thr Glu Gln Ile Amn Ala Ala Ile 1 10 15

Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro 20 25 30

Ile Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr
35 40 45

Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val
50 55 60

Val Ser Thr Asn Arg Arg Ile Met Asn Lys Val Pro Ala Ile Thr Ser 65 70 75 80

His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg 85 90 95

His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala 100 105 110

Ala Gln Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys 115 120 125

Glu Pro Asn Leu Ile Ser Val Pro Ala Pro Ile Thr Val Cys Gly Asp 130 140

Ile His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly 145 150 160

Asp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg 165 170 175

Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu 180 185 190

Asn Phe Asn Asp His Phe Trp Leu Leu Arg Gly Asn His Glu Cys Lys 195 200 205

His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu His Lys Tyr 210 215 220

Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro 225 230 235

Leu Ala Ala Leu Met Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile 245 250 255 Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe

Arg Glu Ile Pro Ser His Gly Leu Met Cys Asp Leu Leu Trp Ala Asp

Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu

Asp Ile Val Asn Ser Lys Thr Met Val Pro His His Gly Lys Met Ala

Pro Ser Arg Asp Met Phe Val Pro Asn Ser Val Arg Gly Cys Ser Tyr

Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu 345

Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met

Tyr Lys Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe

Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu 390

Lys Tyr Glu Asn Asn Val Met Asn Ile Arg Gln Phe Asn Met Thr Pro

His Pro Tyr Trp Leu Pro Asp Phe Met Asp Val Phe Thr Trp Ser Leu

Pro Phe Val Gly Glu Lys Val Thr Glu Het Leu Val Ala Ile Leu Asn

Ile Cys Thr Glu Asp Glu Leu Glu Asn Asp Thr Pro Val Ile Glu Glu

Leu Val Gly Thr Asp Lys Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala

Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu 490 485

Asp Asp Glu His Arg Arg 500

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: G4-PCR-A

24

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22: 24 GCCCTATCGT GCACTCACCG ACGC

- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs

 - (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: G4-PCR-B
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GTGAAGGCCC TACTGAGCCA GGAG

IT IS CLAIMED:

- 1. A polypeptide composition comprising a polypeptide effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin.
 - 2. A composition of claim 1, wherein the polypeptide composition contains a calcineurin interacting (CNI) polypeptide.
- 3. A composition of claim 2, wherein the polypeptide composition contains a polypeptide having an amino acid sequence selected from the group consisting of sequences represented by SEQ ID NO:2 and SEQ ID NO:5.
- 4. A composition of claim 2, wherein the polypeptide has an amino acid sequence of between 15 and 915 amino acids in length.
 - A composition of claim 2, wherein the polypeptide composition contains a
 polypeptide having an amino acid sequence comprising the c-terminal 306 amino acids of a CNI
 protein.

20

- 6. An isolated nucleic acid having a sequence encoding a polypeptide of any of claims 1-5.
- A nucleic acid of claim 6, wherein the nucleic acid has a sequence selected from
 the group consisting of nucleic acid sequences represented by SEQ ID NO:3 and SEQ ID
 NO:6.
 - 8. A method of identifying a small molecule immunosuppressant compound, comprising
- 30 constructing a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains an (A) subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI polypeptide,

contacting the cell with a small molecule, and

identifying the small molecule as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins.

9. A method of claim 8, wherein the cell is a yeast cell.

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- 10. A method of claim 8, wherein one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain.
- 10 11. A method of claim 8, wherein the subunit of calcineurin is selected from the group consisting of yeast CNA1 and yeast CNA2.
 - 12. A method of claim 8, wherein the subunit of calcineurin is an "A" subunit of human calcineurin.

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- 13. A method of claim 8, wherein the CNI polypeptide is yeast CNI polypeptide.
- 14. A method of claim 8, wherein the CNI polypeptide is yeast CNIc polypeptide.
- 20 15. A method of claim 8, wherein the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell.
- 16. A yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where said mutation prevents expression of a
 functional calcineurin-interacting polypeptide from said genomic copy.

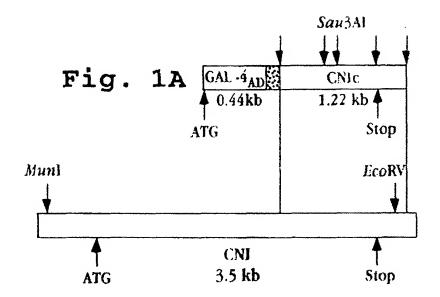
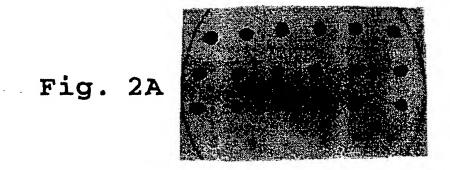
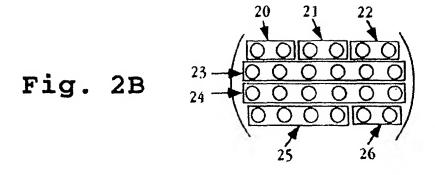


Fig. 1B





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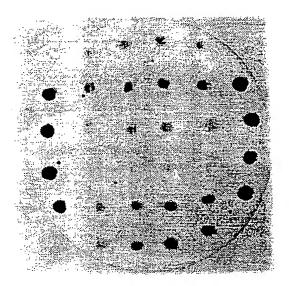


Fig. 3A

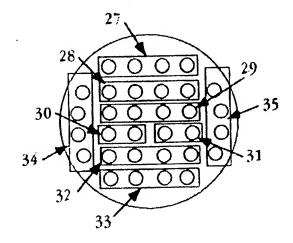
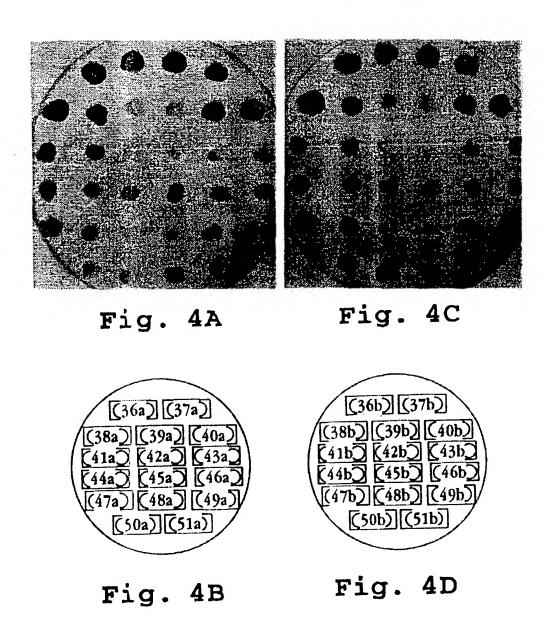
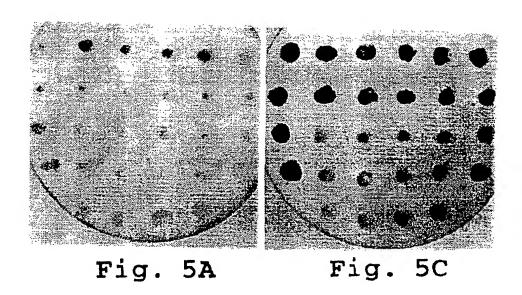
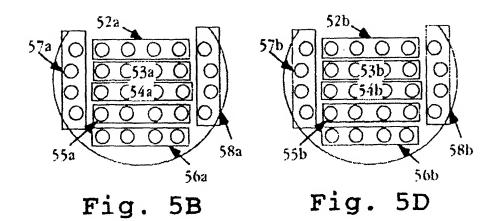


Fig. 3B

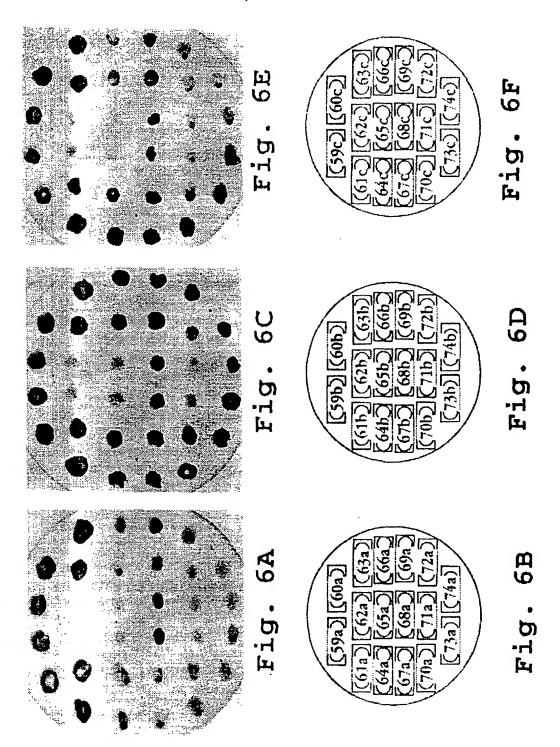
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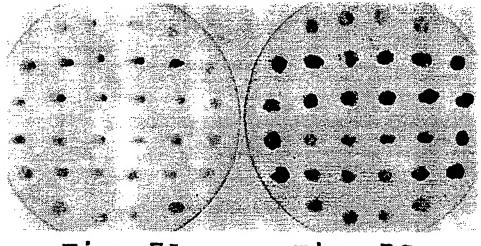


Fig. 7A

Fig. 7C

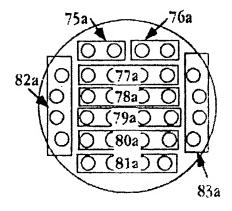


Fig. 7B

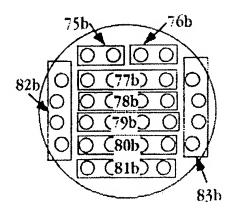


Fig. 7D

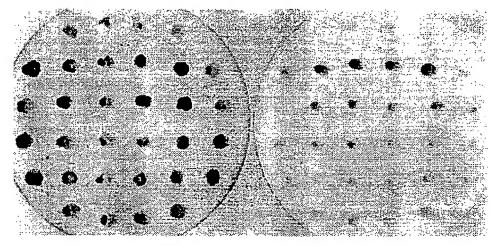


Fig. 7E

Fig. 7G

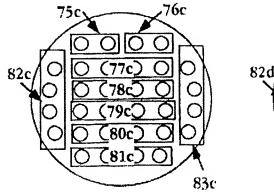


Fig. 7F

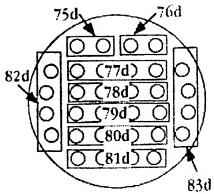
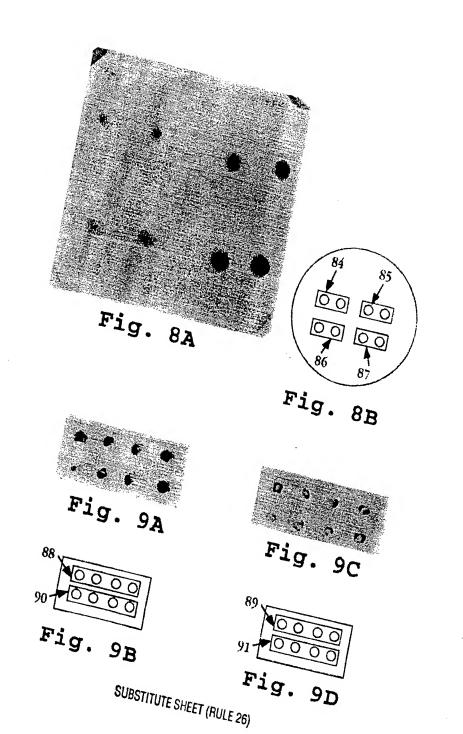
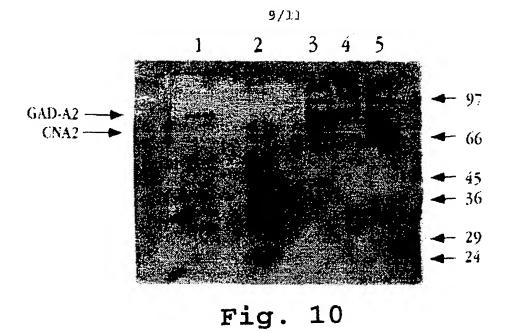


Fig. 7H



8NSDOCID: <WO___9612806A1_1_3

WO 96/12806 PCT/US95/13580



WELLS — CNM1

Fig. 11

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ccgtcccact ggtgcagaca acaacactac aagtctttgt aaagcttgcc gaacccatag 541
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Fig. 12

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Fig. 13

INTERNATIONAL SEARCH REPORT Ternational Application No.

PCT/US 95/13580

A. CLASS	C12N15/31 C07K14/395 G01N33/ C12N1/16 //(C12N1/16,C12R1:86	768 C12N15/62 C	1201/00
According t	to International Patent Classification (IPC) or to both national class	Sistemand IPC	
	S SEARCHED		
IPC 6	ocumentation searched (classification system followed by classific C12N C07K G01N C12Q	ation symbols)	
Documenta	tion searched other than minimum documentation to the extent tha	t such documents are included in the fi	elds scarched
Electronic d	data base consulted during the international search (name of data b	ase and, where practical, search terms	used)
C. DOCUM	MENTS CONSIDERED TO BE RELEVANT		
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х	Swissprot Database entry YKO1_YE Accession number P36117; June O1 Duesterhoeft A. et al.:'Hypothet kD protein in YPT52-GCN3 interge region.'	.,1994 cical 102.5	1-5
Х	Emfun Database entry Scykr021w Accession number Z28246; May 10, Duesterhoeft A. et al.:'S. cerev chromosome XI reading frame ORF	risiae	6,7
P,X	MOLECULAR BIOLOGY OF THE CELL, 5 (SUPPL.). 1994. 141A., HUANG L ET AL 'A novel protein that interacts with calcineurin in vivo' see abstract 818		1,2,6, 8-15
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X Furt	ther documents are listed in the continuation of box C.	Patent (amily members are)	isted in annex.
"A" document defining the general state of the art which is not considered to be of particular relevance. "E" earlier document but published on or after the international filing date. "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another obtation or other special reason (as specified). "O" document referring to an oral disclosure, use, exhibition or other means. "P" document published prior to the international filing date but		"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention. "X" document of particular relevance; the claimed invention cannot be considered nowd or cannot be considered to involve an inventive step when the document is taken alone. "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "&" document member of the same patent family.	
Date of the actual completion of the international search 21 March 1996		Date of mailing of the international search report 0 2. 04. 96	
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijt Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+ 31-70) 340-3016		Authorized officer Espen, J	

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C.(Continua	DOO) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CURR BIOL, 2 (1). 1992. 18-20., CYERT M S 'IMMUNOSUPPRESSANTS HIT THE TARGET' see the whole document	

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